

Time-dependent changes in the microenvironment of injured spinal cord affects the therapeutic potential of neural stem cell transplantation for spinal cord injury

Nishimura *et al.*



RESEARCH

Open Access

Time-dependent changes in the microenvironment of injured spinal cord affects the therapeutic potential of neural stem cell transplantation for spinal cord injury

Soraya Nishimura^{1,2}, Akimasa Yasuda¹, Hiroki Iwai^{1,2}, Morito Takano^{1,2}, Yoshiomi Kobayashi^{1,2}, Satoshi Nori¹, Osahiko Tsuji³, Kanehiro Fujiyoshi⁴, Hayao Ebise⁵, Yoshiaki Toyama¹, Hideyuki Okano^{2*} and Masaya Nakamura^{1*}

Abstract

Background: The transplantation of neural stem/progenitor cells (NS/PCs) at the sub-acute phase of spinal cord injury, but not at the chronic phase, can promote functional recovery. However, the reasons for this difference and whether it involves the survival and/or fate of grafted cells under these two conditions remain unclear. To address this question, NS/PC transplantation was performed after contusive spinal cord injury in adult mice at the sub-acute and chronic phases.

Results: Quantitative analyses using bio-imaging, which can noninvasively detect surviving grafted cells in living animals, revealed no significant difference in the survival rate of grafted cells between the sub-acute and chronic transplantation groups. Additionally, immunohistology revealed no significant difference in the differentiation phenotypes of grafted cells between the two groups. Microarray analysis revealed no significant differences in the expression of genes encoding inflammatory cytokines or growth factors, which affect the survival and/or fate of grafted cells, in the injured spinal cord between the sub-acute and chronic phases. By contrast, the distribution of chronically grafted NS/PCs was restricted compared to NS/PCs grafted at the sub-acute phase because a more prominent glial scar located around the lesion epicenter enclosed the grafted cells. Furthermore, microarray and histological analysis revealed that the infiltration of macrophages, especially M2 macrophages, which have anti-inflammatory role, was significantly higher at the sub-acute phase than the chronic phase. Ultimately, NS/PCs that were transplanted in the sub-acute phase, but not the chronic phase, promoted functional recovery compared with the vehicle control group.

Conclusions: The extent of glial scar formation and the characteristics of inflammation is the most remarkable difference in the injured spinal cord microenvironment between the sub-acute and chronic phases. To achieve functional recovery by NS/PC transplantation in cases at the chronic phase, modification of the microenvironment of the injured spinal cord focusing on glial scar formation and inflammatory phenotype should be considered.

Keywords: Spinal cord injury, Neural stem/progenitor cells, Cell transplantation, Chronic phase, Microenvironment

* Correspondence: hidokano@a2.keio.jp; masa@a8.keio.jp

²Department of Physiology, Keio University School of Medicine, 35 Shinanomachi, Shinjuku, Tokyo 160-8582, Japan

¹Department of Orthopaedic Surgery, 35 Shinanomachi, Shinjuku, Tokyo 160-8582, Japan

Full list of author information is available at the end of the article

Background

The injured spinal cord exhibits little spontaneous recovery and, as a result, many spinal cord injury (SCI) patients suffer from permanent functional impairments, such as motor and sensory dysfunction, and bladder and rectal disturbance. However, some previous reports have shown that neural stem/progenitor cells (NS/PCs) transplanted into the injured spinal cord of rodents [1-6] and non-human primates [7], 7–10 days post-injury (DPI), promote functional recovery after SCI. These reports indicate that NS/PC transplantation has therapeutic potential for SCI when performed during the sub-acute phase. However, patients continue to seek new therapies for SCI many years after their original injury, and most are therefore in the chronic phase. Although many researchers have sought to achieve functional recovery at the chronic phase of SCI by NS/PC transplantation, with one exception [8], no significant recovery of motor function has been obtained in animal models of chronic-phase SCI [9-12]. Despite differences in the survival rate, the cell types derived from the grafted NS/PCs and the distribution of grafted cells transplanted at the sub-acute versus the chronic phase remain unknown. Thus, it remains unanswered as to why grafted NS/PCs do not exert therapeutic benefits in the injured spinal cord at the chronic phase. To address this question, this study analyzed fetus-derived NS/PCs transplanted into the injured spinal cord of mice at 9 DPI and 42 DPI.

To assess the survival rate of grafted cells, we performed quantitative analysis using bioluminescence imaging (BLI) on a weekly basis until 42 days after transplantation. BLI is a powerful tool for the detection of exclusively living grafted cells that stably express luciferase in living animals after administration of luciferin, the luciferase substrate, because the luciferin-luciferase reaction depends on oxygen and ATP [13]. In this study, no significant difference in the survival rate of grafted cells between the sub-acute and chronic transplanted (TP) groups was observed at each experimental time point. Immunohistology also revealed no significant difference in the differentiation pattern of grafted NS/PCs between the two groups. In addition, inflammatory cytokines and growth factors, which influence the survival rate and differentiation characteristics of grafted cells, were expressed at similar levels at both phases. By contrast, the grafted cells were distributed broadly from the epicenter to rostral and caudal sites in the sub-acute TP group, whereas they remained near the lesion epicenter, due to extensive glial scarring, in the chronic TP group. Moreover, prominent macrophages distributed at and around the lesion epicenter in the sub-acute phase by immunohistochemistry, and microarray analysis demonstrated that the expression of Arginase-1, which is associated with M2 macrophages,

was up-regulated significantly at the sub-acute phase than the chronic phase. These findings indicated that the characteristics of post-SCI inflammation are different between the sub-acute and chronic phases.

Consequently, the grafted NS/PCs did not promote any motor functional or histological recovery in the chronic TP group, while the sub-acute TP group demonstrated significant recovery compared with the vehicle control group. Taken together, these data suggest that a combination therapy of NS/PC transplantation with control of glial scar formation or inflammatory reaction may be critical to achieving functional recovery for chronic SCI.

Results

In vitro characterization of transgenic mouse-derived NS/PCs that ubiquitously express fluorescent protein-fused luciferase (fLuc)-cp156

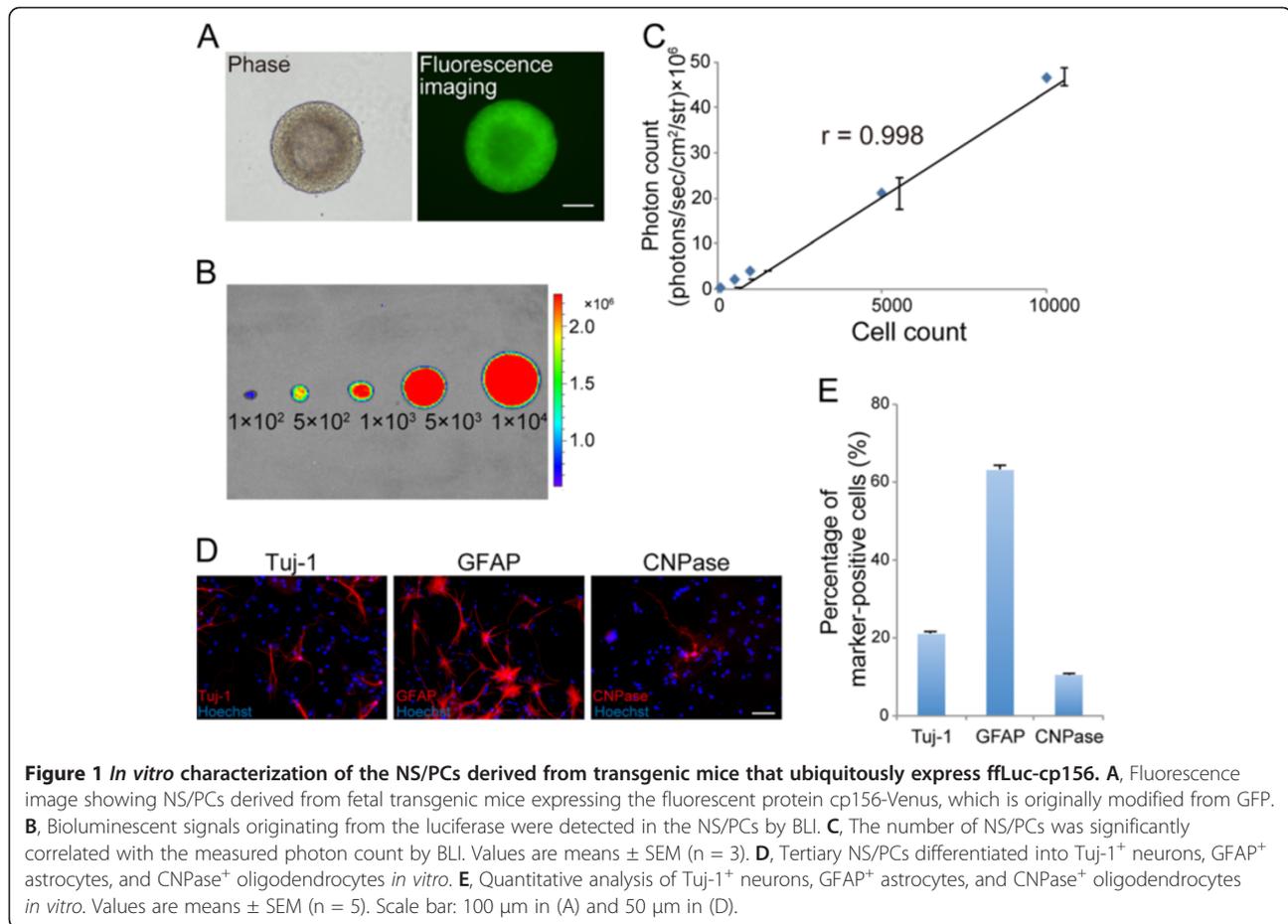
To identify and monitor the grafted cells by bio-imaging, a transgenic mouse that ubiquitously expresses fLuc-cp156 was previously developed [14]. NS/PCs derived from this transgenic mouse showed strong and stable emission of fLuc-cp156 *in vitro* (Figure 1A, B). The number of NS/PCs and the photon counts measured by BLI were significantly correlated (Figure 1B, C).

We proceeded to perform differentiation and proliferation assays of these NS/PCs *in vitro*. The NS/PCs differentiated into β III tubulin (Tuj-1)⁺ neurons ($21.0 \pm 0.5\%$), glial fibrillary acidic protein (GFAP)⁺ astrocytes ($63.0 \pm 1.5\%$), and 2'3'-cyclic nucleotide 3'-phosphodiesterase (CNPase)⁺ oligodendrocytes ($10.4 \pm 0.5\%$) *in vitro* (Figure 1D, E). ATP production, an indirect measurement, was used to assess NS/PC proliferative ability [15]. The doubling time of the NS/PCs was 28.8 ± 0.8 h. The differentiation rate and proliferative ability of NS/PCs obtained from the transgenic mice were equivalent to those previously reported for wild-type NS/PCs [16,17].

Comparison of the injured spinal cord microenvironment between the sub-acute and chronic phases

To clarify differences in the microenvironment of the injured spinal cord between the sub-acute and chronic phases, histological analyses of spinal cord tissues at 9 DPI and 42 DPI were performed. Spinal cord atrophy and glial scar formation were more prominent at and around the lesion epicenter at 42 DPI than at 9 DPI (Figure 2A). A significantly larger CS56⁺ chondroitin sulfate proteoglycan (CSPG) area was detected at the lesion site at 42 DPI than at 9 DPI (Figure 2A, B). Furthermore, Iba1⁺ macrophages infiltrated area was more prominent at the lesion site at 9 DPI but not at 42 DPI (Figure 2A, C).

To analyze the gene expression profile in the injured spinal cord, we performed microarray analysis



to provide a global analysis of the gene expression profile of spinal cord tissues at 9 DPI and 42 DPI. As a control, samples of uninjured naïve spinal cord were prepared. Principal component analysis (PCA) of all the microarray data revealed that the samples of the intact, 9 DPI and 42 DPI groups were clustered at different locations (Figure 2D). Hierarchical clustering of the target genes, which were narrowed down by cut-off values for expression levels and by fold change, revealed that the gene expression profiles of both injured groups were dramatically different from that of the intact group. Furthermore, the gene expression pattern at 9 DPI was similar to that at 42 DPI, but the magnitude of changes in gene expression differed between the two injury groups. At 9 DPI, the magnitude of gene expression changed more remarkably than that at 42 DPI (Figure 2E).

Subsequently, we focused on individual gene expression levels. No significant differences were observed in the expression levels of genes for individual inflammatory cytokines or growth factors between the two injury groups (Figure 2F, G). By contrast, the expression of CD36 and CD68, which are expressed on monocytes and macrophages, was significantly elevated at 9 DPI

(Figure 2H). These results were consistent with the immunostaining results for Iba1 (Figure 2A, C). Interestingly, the expression of arginase-1, which is associated with anti-inflammatory M2 macrophages, was also significantly higher at 9 DPI than at 42 DPI, whereas no significant differences were observed in the expression levels of genes associated with pro-inflammatory M1 macrophages, such as CD86 or inducible nitric oxide synthase (iNOS). Consistent with these findings, immunohistological analysis revealed more Iba1 and arginase-1 double-positive cells at 9 DPI ($14.1 \pm 1.5\%$) than at 42 DPI ($0.9 \pm 0.4\%$) (Figure 2I, J).

We also evaluated the phagocytic activity of the macrophages recruited into the injured spinal cord by performing immunohistochemistry for LAMP2, a marker for endosomes or lysosomes. At 9 DPI, substantial numbers of infiltrating Iba1, arginase-1, and LAMP2 triple-positive cells were observed at and near the lesion epicenter. In contrast, only a small amount of LAMP2-positive phagocytes were localized to the lesion epicenter at 42 DPI (Figure 2I). In addition, no significant expression of myeloperoxidase, a marker of neutrophils, was observed at either 9 DPI or 42 DPI (Figure 2H).

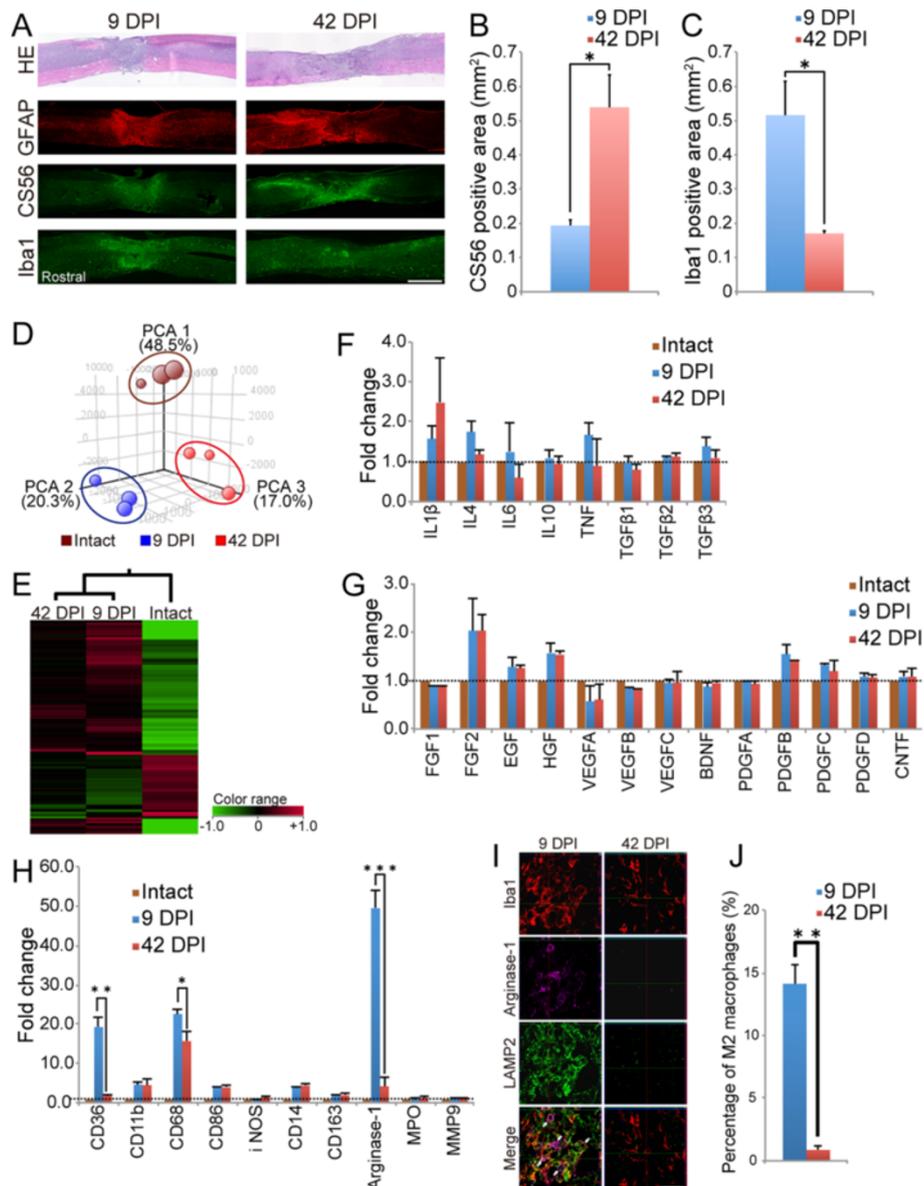


Figure 2 Comparison of the microenvironment of the injured spinal cord at 9 DPI versus 42 DPI. **A**, Representative images of HE staining and immunofluorescence staining for GFAP, CS56, and Iba1 in sagittal sections. **B**, CSPGs accumulation was more prominent at the lesion site at 42 DPI than at 9 DPI. Values are means \pm SEM ($n = 3$). $^*P < 0.05$. **C**, More Iba1-positive cells were distributed at the lesion site at 9 DPI than at 42 DPI. Values are means \pm SEM ($n = 3$). $^*P < 0.05$. **D**, Overview of all the microarray data by PCA. The samples of each group were clustered at different locations on 3D visualization. **E**, Hierarchical clustering analysis showed that the gene expression pattern at 9 DPI was similar to that at 42 DPI. However, the magnitude of changes in gene expression differed between the two injury groups. Green tiles show downregulated genes and red tiles indicate upregulated genes. **F-H**, Gene expression signals of cytokines (**F**), growth factors (**G**), and markers of inflammatory cells (**H**) at 9 DPI and 42 DPI. The gene expression levels of markers associated with microglia/macrophages significantly differed between 9 DPI and 42 DPI, but those of all cytokines and growth factors did not. Data are the mean fold-change values versus intact samples. Values are means \pm SEM ($n = 3$). $^*P < 0.05$, $^{**}P < 0.01$, $^{***}P < 0.001$. **I**, Representative images of immunofluorescence staining for Iba1, arginase-1, and LAMP2 in sagittal sections of the lesion epicenter at 9 DPI and 42 DPI. Arrows: Iba1⁺/arginase-1⁺/LAMP2⁺ triple-positive cells. **J**, At the lesion epicenter, more Iba1⁺/arginase-1⁺ M2 macrophages had infiltrated at 9 DPI compared with 42 DPI. Values are means \pm SEM ($n = 3$). $^{**}P < 0.01$. Scale bar: 1000 μ m in (**A**) and 50 μ m in (**J**).

Comparison of survival rates and differentiation phenotypes of the grafted NS/PCs between the sub-acute and chronic TP groups

BLI analysis only detects luminescent photon signals from living cells, and the number of NS/PCs and the photon count *in vitro* were significantly correlated. To investigate whether similar correlativity was observed *in vivo*, various numbers of NS/PCs (approximate range 2.5×10^4 to 5×10^5 cells) were transplanted into the intact spinal cord of mice (Figure 3A). These

data revealed that the photon count was significantly proportional to the number of grafted cells *in vivo* (Figure 3B).

We next analyzed the survival rate of the grafted cells on a weekly basis until 42 days after transplantation using BLI (Figure 3C). At 7 days after transplantation, BLI analysis revealed that the survival of grafted cells was reduced to $12.4 \pm 5.6\%$ in the sub-acute TP group and to almost the same level in the chronic TP group ($15.2 \pm 2.9\%$). At 42 days after

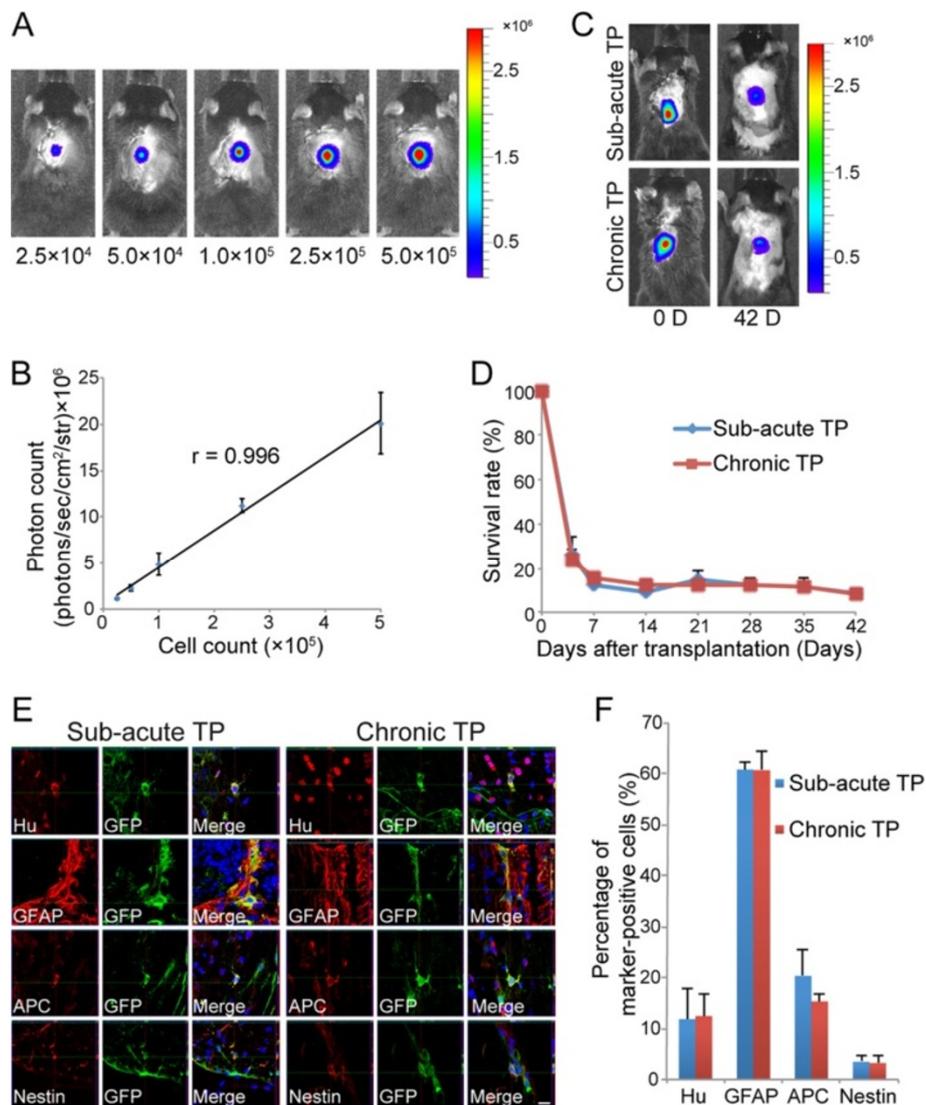


Figure 3 Survival rate and differentiation phenotype of grafted cells in the sub-acute and chronic TP groups. **A**, Representative *in vivo* BLI of naïve mice transplanted with various numbers of NS/PCs. **B**, The number of NS/PCs and the measured photon count were significantly correlated *in vivo*. Values are means \pm SEM ($n = 3$). **C**, Representative *in vivo* BLI of the sub-acute and chronic TP groups immediately and 42 days after transplantation. **D**, Quantitative analysis using BLI revealed no significant difference in the survival rate of grafted cells between the sub-acute and chronic TP groups up to 42 days after transplantation. Values are means \pm SEM ($n = 10$). **E**, GFP⁺ grafted cells differentiated into Hu⁺ neurons, GFAP⁺ astrocytes, and APC⁺ oligodendrocytes in the sub-acute and chronic TP groups. **F**, The differentiation rate of grafted cells into the three neural cell lineages did not significantly differ between the sub-acute and chronic TP groups 42 days after transplantation. Only a small number of nestin⁺ neural progenitor cells was detected in the sub-acute and chronic TP groups. Values are means \pm SEM ($n = 3$). Scale bar: 10 μ m in (C).

transplantation, approximately 8% of the cells survived in both the sub-acute and chronic groups ($8.6 \pm 2.6\%$ vs. $8.3 \pm 1.9\%$). The survival rate of grafted cells did not significantly differ between the sub-acute and chronic TP groups at any time point examined (Figure 3D).

To evaluate the differentiation phenotype of the grafted cells *in vivo*, immunostaining for cell markers was performed 42 days after transplantation for both the sub-acute and chronic groups. Green fluorescent protein (GFP)⁺ grafted cells differentiated into all three neural lineages in both groups (Figure 3E). Quantitative analyses revealed that in the sub-acute and chronic TP groups, most of the grafted cells differentiated into GFAP⁺ astrocytes ($60.8 \pm 1.6\%$ and $60.7 \pm 3.7\%$), followed by adenomatous polyposis coli antigen (APC)⁺ oligodendrocytes ($20.3 \pm 5.1\%$ and $15.3 \pm 1.3\%$) and Hu⁺ neurons ($11.8 \pm 6.0\%$ and $12.4 \pm 4.2\%$). The differentiation rates of neurons, astrocytes, and oligodendrocytes did not significantly differ between the sub-acute and chronic TP groups (Figure 3F). Nestin-positive cells represented around 3% of the grafted cells in both TP groups.

Grafted cells were limited to the lesion epicenter in the chronic TP group due to extensive glial scar formation

To examine whether the prominent glial scarring seen in the chronic TP group affected the distribution of grafted

cells, we performed immunostaining for GFP in both TP groups. In sagittal sections of the sub-acute TP group, GFP⁺ grafted cells were found at the epicenter and at rostral and caudal sites, whereas in the chronic TP group, grafted cells were seen almost exclusively at the lesion epicenter (Figure 4A, B). Quantitative analysis of the GFP⁺ areas in the axial sections revealed that the GFP⁺ area in the sub-acute TP group, which spread from 3 mm rostral to 2 mm caudal from the lesion epicenter, was significantly larger than that in the chronic TP group (Figure 4C). Moreover, to clarify what causes the difference of distribution of grafted cells, double immunostaining for GFP and CS56 was performed in both TP groups. The GFP⁺ grafted cells were surrounded and enclosed by CS56⁺ CSPG areas at the lesion site in the chronic TP group, whereas the grafted cells distributed beyond CSPGs areas due to less accumulation of CSPG around the lesion site in the sub-acute TP group (Figure 4D, E).

NS/PC transplantation at the sub-acute phase, but not at the chronic phase, contributes to the preservation and/or enhancement of myelination and axonal growth

To compare the effects of NS/PC transplantation on the injured spinal cord between the sub-acute and chronic groups, axial sections were examined histologically using hematoxylin-eosin (HE) staining (Figure 5A). In sections

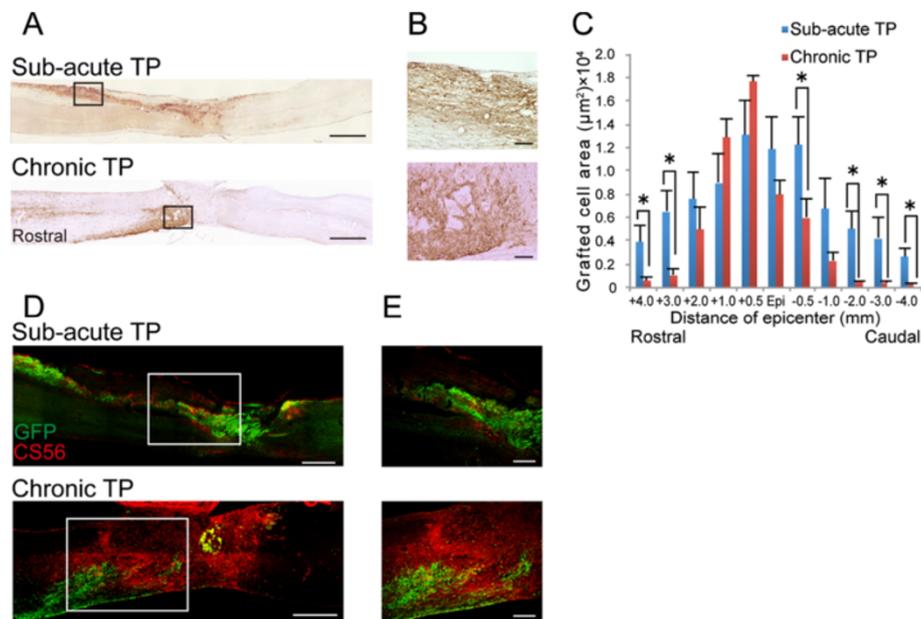


Figure 4 Distribution of grafted cells in the sub-acute and chronic TP groups. **A**, Representative images of GFP-immunostained sagittal sections in the sub-acute and chronic TP groups 42 days after transplantation. **B**, Higher-magnification images of the boxed areas in **(A)**. **C**, Quantitative analysis of the GFP⁺ grafted cell area in axial sections 42 days after transplantation. Grafted cells were located at the epicenter, rostral, and caudal sites in the sub-acute TP group, whereas they were limited to the lesion epicenter in the chronic TP group. Values are means \pm SEM (n = 4). *P < 0.05. **D**, Representative images of GFP- and CS56-immunostained sagittal sections from the sub-acute and chronic TP groups. **E**, Higher-magnification images of the boxed area in **(D)**. In the sub-acute TP group, the GFP⁺ grafted cells migrated away from the graft site due to less accumulation of CS56⁺ CSPG around the lesion site, while the robust CSPGs prevented further migration of the grafted cells in the chronic TP group. Scale bar: 1000 μ m in **(A)**, 100 μ m in **(B)**, 500 μ m in **(D)** and 200 μ m in **(E)**.

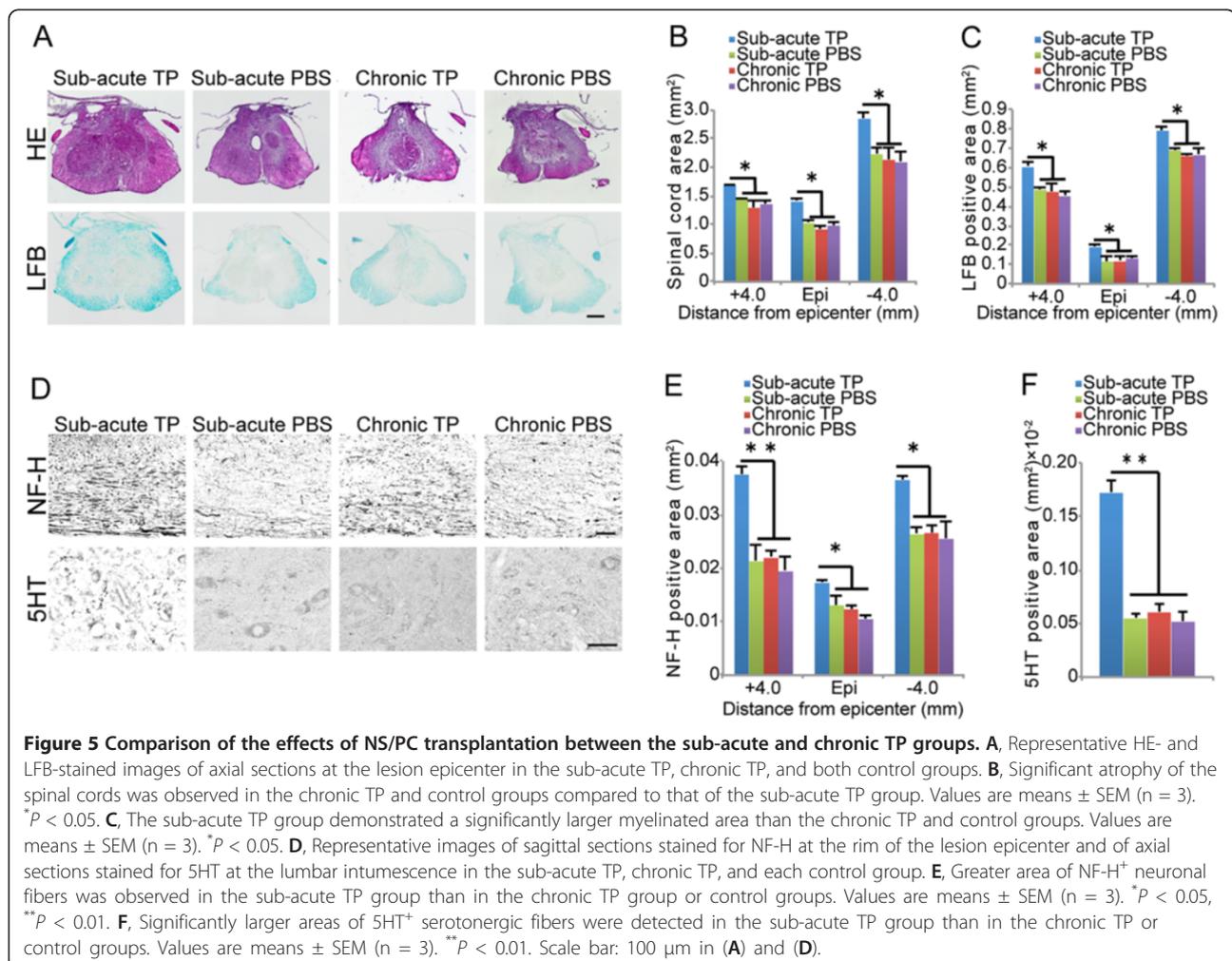
at the lesion epicenter and 4 mm rostral and caudal to it, the HE-stained images revealed significant atrophy of the spinal cord in the sub-acute phosphate-buffered saline (PBS) group compared to the sub-acute TP group. However, the experimental transverse area of the spinal cord did not significantly differ between the chronic TP and PBS groups (Figure 5B). In addition, the sub-acute TP group demonstrated a significantly larger myelinated area compared to the sub-acute PBS group at the lesion epicenter and in sections 4 mm rostral and caudal to the epicenter (Figure 5A), whereas the myelinated area did not significantly differ between the chronic TP and PBS groups (Figure 5C).

Next, to investigate axonal growth after NS/PC transplantation, immunostaining for 200 kDa neurofilament (NF-H) and 5-hydroxytryptamine (5HT) was performed in all experimental groups (Figure 5D). The NF-H⁺ areas at the lesion site as well 4 mm rostral and caudal to it significantly differed between the sub-acute TP group and the other groups (Figure 5E). Furthermore, the sub-acute TP group demonstrated a significantly larger area

of 5HT⁺ serotonergic fibers in the distal cord compared to the other three groups (Figure 5F). By contrast, the chronic TP group demonstrated no significant differences in the NF-H⁺ and 5HT⁺ areas compared to the chronic PBS group.

NS/PC transplantation at the sub-acute phase, but not at the chronic phase, promotes motor function and electrophysiological recovery after SCI

Finally, we evaluated locomotor functional recovery by Basso Mouse Scale (BMS) score, Rotarod testing, and DigiGait. We confirmed that the mice exhibited complete paralysis of the hindlimbs by a BMS score of 0 at 1 DPI. At 7 DPI, the hind limb locomotor functions had recovered spontaneously to an approximate BMS score of 2, and plateaued at an approximate BMS score of 3, in all experimental groups except the sub-acute TP group. In the sub-acute TP group, significantly greater functional recovery was observed compared to the sub-acute PBS group at 14 DPI and thereafter. By contrast, the chronic TP group did not show any



functional recovery, with BMS scores comparable to those of the chronic PBS group (Figure 6A).

The gait performance of the mice in each group was analyzed using the DigiGait system at 42 days after cell transplantation or PBS injection. In both PBS control groups and the chronic TP group, a subset of experimental animals (2 of 10 mice in each group) were unable to walk sufficiently well on a treadmill moving at 7 cm/s to perform the test, but all the mice in the sub-acute TP group could perform the test. Gait analysis revealed a significantly longer stride length in the sub-acute TP group than in the sub-acute PBS group, whereas no significant difference in stride length was detected between the chronic TP and either PBS group (Figure 6B). Consistent with these findings, in the Rotarod test at 42 days after cell transplantation or PBS injection, the mice in the sub-acute TP group remained on the rotating rod for a significantly longer time than those of the sub-acute PBS group, but no significant difference was observed between the chronic TP group and either PBS group (Figure 6C).

Electrophysiological examinations performed at 42 days after cell transplantation or PBS injection by motor-evoked potential (MEP). MEP waves were detected in all mice of the sub-acute TP group, and the average signal-to-response latency was 6.1 ± 0.2 ms and the average first wave amplitude was 134.0 ± 31.5 mv. However no MEP waves were observed at all in 5 out of the 7 mice in the chronic TP group and at all mice in the sub-acute and chronic PBS groups (Figure 6D). Taking the results of all the motor functional analyses together, no functional recovery of the hindlimbs was observed in the chronic TP group.

Discussion

Previous studies have indicated that NS/PC transplantation at the sub-acute phase has therapeutic potential for SCI [1-7,18]. At the chronic phase, however, NS/PC transplantation does not result in functional recovery. Many researchers have sought to improve motor function in chronic SCI by NS/PC transplantation. However,

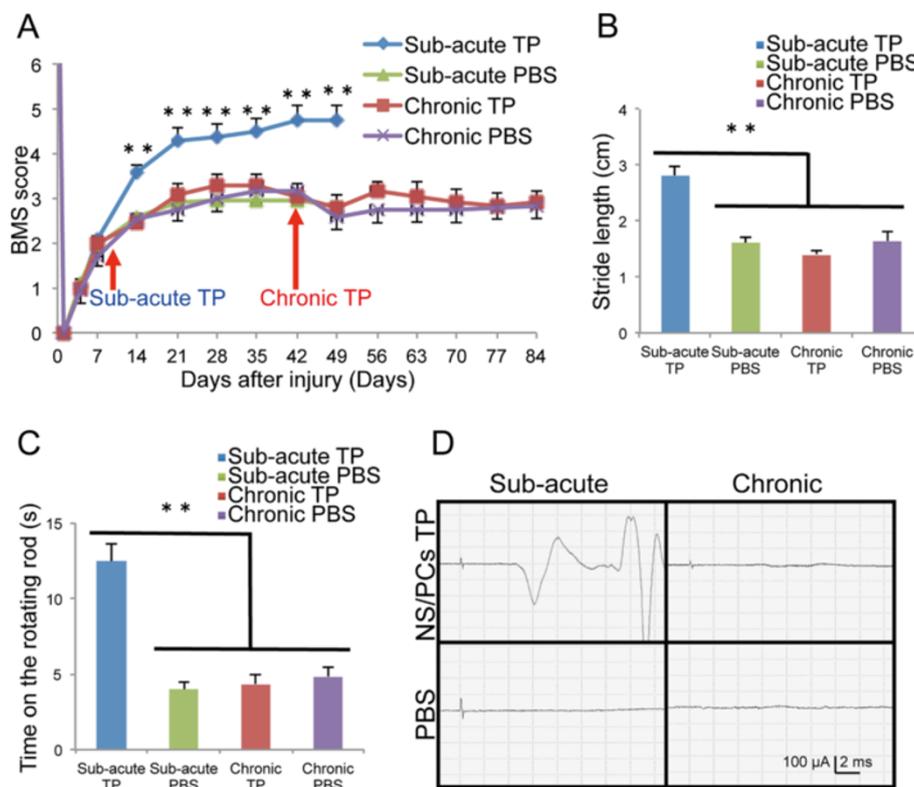


Figure 6 Motor function and electrophysiological recovery after NS/PC transplantation. **A**, Hindlimb motor function was assessed weekly by BMS score. The sub-acute TP group exhibited significantly better functional recovery than the vehicle control group at 14 DPI and thereafter, whereas no significant difference was observed between the chronic TP group and the vehicle control group. Values are means \pm SEM ($n = 10$). $**P < 0.01$. **B**, Treadmill gait analysis using the DigiGait system revealed a significantly longer stride in the sub-acute TP group than in the chronic TP group or either control group 42 days after cell transplantation or PBS injection. Values are means \pm SEM (sub-acute TP $n = 10$, chronic TP and each control $n = 8$). $**P < 0.01$. **C**, The sub-acute TP group walked for a significantly longer time on the rotating rod than the chronic TP group or either control group 42 days after cell transplantation or PBS injection. Values are means \pm SEM ($n = 10$). $**P < 0.01$. **D**, Representative profiles of MEP in the sub-acute TP, chronic TP, and both control groups 42 days after cell transplantation or PBS injection.

most studies obtained no significant functional recovery, and no consensus exists concerning the survival and/or fate of grafted cells in chronic-phase transplantation [9-12]. For example, some reports demonstrated that cells grafted at the chronic phase of SCI have a poor survival rate [9,12], while Cusimano et al. reported no significant difference in survival rate between NS/PC transplantation at the sub-acute versus the chronic phase [11]. However, these findings were based on histological examinations only. To clarify this issue, we performed sub-acute- and chronic-phase transplantations of NS/PCs derived from transgenic mice that ubiquitously express fLuc-cp156 on a C57BL/6J background [14] to treat SCI in adult mice. These NS/PCs showed strong bioluminescent and fluorescent signals originating from fLuc-cp156 [14], enabling the easy detection of surviving grafted cells in living animals noninvasively by BLI [4,13] and confirmation of integrated cells within the injured spinal cord by immunohistology for GFP without using a lentiviral vector.

Using cells from these mice, we demonstrated that the NS/PCs grafted during the chronic phase of SCI had a similar survival rate to those transplanted at the sub-acute phase. We also observed no significant difference between the two groups in the differentiation phenotypes of the grafted cells. The survival and/or differentiation potential of grafted cells can change remarkably due to microenvironmental changes in the injured spinal cord [19]. A previous study showed that grafted NS/PCs mainly differentiate into astrocytes after transplantation at the acute phase [4]. Inflammatory cytokines, such as IL-1 β , IL-6, and TNF- α , also increase dramatically in the injured spinal cord during the acute phase [20,21], and these cytokines induce NS/PCs to differentiate into astrocytes. Moreover, growth factors affect the fate of grafted NS/PCs. For example, EGF, FGF, NT3, and PDGF have demonstrated beneficial effects on the survival of NS/PCs [10,12,22]. HGF and PDGF promote neuronal differentiation [23,24], whereas CNTF enhances glial differentiation *in vitro* [25]. To assess microenvironmental changes in the injured spinal cord, we performed a microarray analysis to exhaustively compare the gene expression profiles between the sub-acute and chronic phases of SCI. The expression levels of all cytokines and growth factors did not significantly differ, consistent with our BLI and histological results on the survival and/or fate of the grafted cells. These data suggested that the microenvironments of the injured spinal cord were not significantly different at the sub-acute and chronic SCI phases with respect to the expression of factors known to influence the survival rate and differentiation of grafted NS/PCs.

Many reports concern possible mechanisms underlying the therapeutic effects of NS/PC transplantation

for SCI; for example, grafted cells contribute to the reconstruction of neural circuits and remyelination [2,6,17,26]. NS/PCs also secrete neurotrophic factors, which support the survival of host neural cells and enhance axonal growth and angiogenesis [27-30]. In this study, motor function recovery was observed in the sub-acute TP group, but not in the chronic TP group. Consistent with this result, more NF-H⁺ neuronal fibers and 5HT⁺ serotonergic fibers, which are involved in motor function recovery [30-32], were observed in the sub-acute TP group than in the sub-acute PBS group. Moreover, Luxol Fast Blue (LFB) staining revealed significantly larger myelinated areas in the sub-acute TP group than in the sub-acute PBS group. By contrast, no significant differences in NF-H⁺ neuronal fibers, 5HT⁺ serotonergic fibers, or LFB⁺ myelinated areas were observed between the chronic TP and chronic PBS groups. Furthermore, no MEP waves were detected in most mice in the chronic TP group.

As a possible reason for these findings, we refer to the poor migration of grafted cells in the chronic TP group. Since glial scar formation prevented grafted NS/PCs from migrating away from the graft site and integrating with the host spinal cord, the regeneration of neural circuits, remyelination, and/or preservation of host neural cells did not occur in areas rostral and caudal to the injured spinal cord of the chronic TP group. Consistent with this scenario, many previous reports indicate that insufficient migration of grafted cells results in poor functional recovery after SCI [33,34].

Our microarray and histological analyses suggested that more macrophages or microglial cells, including a substantial number of M2 macrophages, were distributed around the lesion epicenter at the sub-acute phase than at the chronic phase. In agreement with this finding, previous studies have reported that infiltration of macrophages and microglial cells peaks at approximately 7 DPI [35,36]. Moreover, M2 macrophages have an anti-inflammatory role and promote axonal growth after SCI [37-39], and their phagocytic activity was shown to contribute to tissue repair in the injured CNS by reducing myelin debris [40-42]. In addition, Busch et al. demonstrated that activated macrophages induced axonal dieback in the injured CNS [43], which was prevented by switching the infiltrating macrophages from M1 to M2 phenotype [39]. Furthermore, grafted NS/PCs also exert immunomodulatory effects through interactions with infiltrating immune cells [44,45]. Especially in the sub-acute phase, grafted NS/PCs can affect the properties of infiltrating macrophages, inducing their shift from M1 to M2 in the injured spinal cord [11,39]. Therefore, NS/PCs transplanted at the sub-acute stage may alter the microenvironment to favor M2, resulting in a synergistic effect that supports the increase in neuronal fibers and

functional recovery. These findings indicate that a novel immunomodulatory strategy, such as the administration of a mediator of phenotypic switching from M1 to M2 or the combined transplantation of NS/PCs and M2 macrophages, may have therapeutic potential in chronic SCI. In contrast, previous studies reported that M1 macrophages impairs recovery of SCI through the production of oxidative metabolites or pro-inflammatory cytokines including TNF- α , which induced the apoptotic cell death of neural precursor cells [42,46,47]. However, the expressions of genes associated with M1 macrophages showed no significant difference among the sub-acute and chronic phases. These findings are consistent with the similar survival rate of the grafted NS/PCs in both TP groups.

Taken together, these results suggest that during the chronic phase of SCI, the injured spinal cord micro-environment appears to be unfavorable for the therapeutic mechanisms of NS/PC transplantation, owing to extensive glial scarring [48] and/or the phenotype of infiltrating macrophages. Accordingly, to improve the therapeutic potential of NS/PC transplantation performed at the chronic phase, altering the microenvironment in the injured spinal cord is likely to be important. For example, suppression of axonal growth inhibitors may improve the microenvironment. CSPGs, which are the most prevalent axonal growth inhibitors, are produced by reactive astrocytes and involved in glial scarring [48]. Chondroitinase ABC (ChABC), which digests CSPG, promotes axonal growth and the migration of grafted NS/PCs [49,50]. Karimi-Abdolrezaee et al. described that CSPGs hinder the survival of grafted cells and the combined therapy of ChABC administration and NS/PCs transplantation promotes the migration and integration of grafted cells [51]. While our data demonstrated that CSPGs prevent the migration of grafted cells, but show no influence on their survival, we have great hopes that a combination of NS/PC transplantation and the administration of a suppressor of axonal growth inhibitor may be effective for inducing functional recovery in chronic SCI.

Conclusions

Microenvironmental changes in SCI affect the survival and/or fate of NS/PCs grafted into the injured spinal cord. The survival rate and differentiation phenotype of grafted cells were similar between chronic-phase and sub-acute-phase transplantation, with no significant difference in the secretion of cytokines and growth factors in the host environment between the two phases. However, the distribution of grafted cells was restricted by the robust glial scar that formed around the lesion epicenter at the chronic phase. Consequently, no functional recovery was observed in this phase. Additionally, in the

sub-acute phase, M2 macrophages, which infiltrated predominantly in this phase, might contribute to the functional repair upon NS/PC transplantation. Alteration of the microenvironment in SCI, focusing on the glial scar and the inflammatory phenotype, appears to be the best possible solution to maximize the therapeutic potential of NS/PC transplantation in the chronic phase.

Methods

NS/PC culture and analysis

NS/PCs were cultured and expanded, as previously reported [52]. Briefly, the striata of transgenic mice ubiquitously expressing fLuc-cp156, a fusion protein of firefly luciferase and a circularly permuted Venus protein [14], were dissociated using a fire-polished glass pipette on embryonic day 14. Venus is a fluorescent protein with fast and efficient maturation that was originally engineered from GFP [53], and therefore grafted cells can be detected as fluorescent Venus signals using anti-GFP antibody [17,26]. Dissociated cells were collected by centrifugation and re-suspended in culture medium, which consisted of Dulbecco's modified Eagle medium/F12 (Sigma-Aldrich, St. Louis, MO, USA) supplemented with a previously described hormone mixture [52]. Human recombinant FGF-2 (Peprotech, Rocky Hill, NJ, USA) and EGF (Peprotech) (20 ng/ml each) were added every 2 days. The cells formed floating cell clusters (neurospheres) within 2–3 days. After propagation for 3 passages, the neurospheres were used for *in vitro* BLI, differentiation, and proliferation assays, and for cell transplantation.

For differentiation analysis, the neurospheres were plated onto poly-L-ornithine/fibronectin-coated 8-well chamber slides (Iwaki; Asahi Glass Co Ltd., Tokyo, Japan) at a density of 1×10^5 cells/ml and cultured in culture medium without serum or growth factors at 37°C in 5% CO₂ and 95% air for 7 days. The differentiated cells were then fixed with 4% paraformaldehyde in 0.1 M PBS and stained with the following primary antibodies for immunocytochemistry: anti-Tuj-1 (mouse IgG, 1:1000, Sigma-Aldrich), anti-GFAP (rat IgG, 1:1000, Invitrogen, Carlsbad, CA, USA), and anti-CNPase (mouse IgG1, 1:1000, Sigma-Aldrich). Nuclei were stained with Hoechst 33258 (10 μ g/ml, Sigma-Aldrich). *In vitro* images were obtained using a fluorescence microscope (BZ-9000; Keyence Co., Osaka, Japan).

The proliferation assay was performed by measuring ATP, which indirectly reflects the number of viable cells [15,17,54]. In brief, NS/PCs were first incubated in culture medium without serum or growth factors in 48-well cell-culture plates (Corning Inc., Corning, NY, USA) at 37°C in 5% CO₂ and 95% air for 24 or 72 h. D-luciferin was then added to each well, and the luminescent signal was detected immediately using a Xenogen-IVIS spectrum cooled charged-coupled device (CCD) optical macroscopic imaging system (Caliper Life

Sciences, Hopkinton, MA, USA). To determine the population doubling time, the ATP assay was modified, as described elsewhere [15,17,54].

SCI model

Adult female C57BL/6J mice (8–10 weeks old, 18–22 g, $n = 52$; Clea, Tokyo, Japan) were anesthetized with an intraperitoneal (i.p.) injection of ketamine (100 mg/kg) and xylazine (10 mg/kg). After laminectomy at the 10th thoracic spinal vertebra (Th10), the dorsal surface of the dura mater was exposed. Contusive SCI was induced using a commercially available SCI device (IH Impactor, Precision Systems and Instrumentation, Lexington, KY, USA), as previously described [4]. This device creates a reliable contusion injury by rapidly applying a force-defined impact (60 kdyn) with a stainless steel-tipped impactor. All experiments were approved by the ethics committee of Keio University and fully in accordance with the Guide for the Care and Use of Laboratory Animals (National Institutes of Health, Bethesda, MD, USA).

Microarray analysis

The injured mice were anesthetized and transcardially perfused with heparinized saline (5 U/ml) at 9 DPI or 42 DPI ($n = 3$ each). Dissected segments of spinal cord at the Th10 level were rapidly frozen and placed in TRIzol (Invitrogen). Total RNA was isolated using an RNeasy Mini Kit (Qiagen Inc., Hilgen, Germany), in accordance with the manufacturer's instructions. As a control, samples of naïve spinal cord were harvested by the same protocol. For microarray analysis, RNA quality was assessed using a 2100 Bioanalyzer (Agilent Technologies Inc., Santa Clara, CA, USA), and 100 ng of total RNA was reverse transcribed, biotin labeled, and hybridized to a GeneChip[®] Mouse Genome 430 2.0 Array (Affymetrix Inc., Santa Clara, CA, USA). The array was then washed and stained in a Fluidics Station 450, according to the manufacturer's instructions [55,56]. The microarrays were scanned using a GeneChip Scanner 3000 7G, and the raw image files were converted to normalized signal intensity values using the MAS 5.0 algorithm. PCA was carried out with Gene Spring GX software (Agilent Technologies Inc.), using the full set of normalized data. For the clustering analysis, the normalized data were narrowed down by the cut-off values of expression levels (>50) and by fold change (>1.5 , versus the signal of intact spinal cord), and statistical analysis was performed using one-way ANOVA followed by the Tukey-Kramer test ($P < 0.05$). The heat map was visualized with Gene Spring GX.

NS/PC transplantation

Various numbers of NS/PCs (approximate range 2.5×10^4 to 5×10^5 cells/ $2 \mu\text{l}$) were transplanted into

uninjured naïve mice ($n = 3$, each), and NS/PCs (5×10^5 cells/ $2 \mu\text{l}$) were transplanted at 9 DPI (sub-acute TP group, $n = 10$) or 42 DPI (chronic TP group, $n = 10$) as previously reported [4,16,17,26,30,57]. The NS/PCs were transplanted into the lesion epicenter with a glass micropipette at a rate of $1 \mu\text{l}/\text{min}$ using a Hamilton syringe ($25 \mu\text{l}$) and a stereotaxic microinjector (KDS 310, Muromachikikai Co. Ltd., Tokyo, Japan). PBS ($2 \mu\text{l}$) was similarly injected into the lesion epicenter of the control mice at each time point (sub-acute and chronic PBS groups, $n = 10$ each).

Bioluminescence imaging

A Xenogen-IVIS spectrum CCD optical macroscopic imaging system was used for *in vitro* and *in vivo* BLI as previously reported [4,16,17]. In brief, the signal intensity of NS/PCs *in vitro* was assessed using plated cells at various cell numbers (approximate range 1×10^2 to 1×10^4 cells/well), and BLI was performed immediately after adding D-luciferin ($150 \mu\text{g}/\text{ml}$) ($n = 3$). The integration time was fixed at 1 min for each image.

In vivo imaging was performed 15 min after the i.p. injection of D-luciferin ($0.3 \text{ mg}/\text{g}$ body weight) with the field-of-view set at 13.2 cm, because the photon count was most stable during this period. The intensity peaked between 10 and 30 min after the i.p. injection of D-luciferin. The integration time was fixed at 5 min for each image. All images were analyzed with Living Image software (Caliper Life Sciences), and the optical signal intensity was expressed as photon flux (photon count) in units of photons/ $\text{s}/\text{cm}^2/\text{steradian}$. Each result was displayed as a pseudo-colored photon count image superimposed on a gray-scale anatomic image. To quantify the measured light, a region of interest was defined in the cell-implanted area, and all values at the same region of interest were examined.

Behavioral analyses

The motor function of each mouse was evaluated weekly using the BMS up to 49 DPI in the sub-acute TP and PBS groups and up to 84 DPI in the chronic TP and PBS groups ($n = 10$ per group) [58]. This assessment was performed by two investigators blinded to the identity of the experimental mice.

Motor coordination was evaluated using a rotating rod apparatus (Rotarod, Muromachikikai Co., Ltd.), which consisted of a plastic rod (3 cm diameter, 8 cm length) with a gritted surface flanked by two large discs (40 cm diameter) ($n = 10$ per group). At 42 days after cell transplantation or PBS injection, each mouse was placed on the rod while it rotated at 20 rpm for 2 min sessions [59]. Three trials were conducted, and the maximum number of seconds the mouse stayed on the rod was recorded.

Gait analysis was performed using the DigiGait system (Mouse Specifics, Quincy, MA, USA) (n = 10 per group) [17,26,60]. Each mouse demonstrated weight-supported hindlimb stepping at 42 days after cell transplantation or PBS injection. The stride length was determined on a treadmill set to a speed of 7 cm/s.

Electrophysiology

Electrophysiological experiments were performed using a Neuropack S1 MEB-9402 (Nihon Kohden, Tokyo, Japan) at 42 days after cell transplantation or PBS injection (n = 7 per group) [26]. The animals were anesthetized with an i.p. injection of ketamine (60 mg/kg) and xylazine (6 mg/kg), and the stimulation was applied through the occipitocervical area of the spinal cord and the hindlimb by needle electrodes. The active electrode was placed in the quadriceps muscle belly, and the reference electrode was placed near the distal quadriceps tendon. The ground electrode was placed on the tail. A stimulus of 0.4 mA intensity, 0.2 ms duration, and 1 Hz interstimulus interval were used. The latency was measured as the length of time from the stimulus to the onset of first response wave. The amplitude was measured from the initiation point of the first response wave to its highest point.

Histological analyses

Injured animals were deeply anesthetized and transcardially perfused with 4% PFA in 0.1 M PBS at 9 DPI or 42 DPI (n = 3 each). The treated animals were similarly prepared 42 days after cell transplantation or PBS injection. The spinal cords were removed, postfixed overnight in 4% PFA, soaked overnight in 10% sucrose, followed by 30% sucrose, embedded in Optimal Cutting Temperature compound (Sakura Finetechnical Co., Ltd., Tokyo, Japan), frozen, and sectioned in the sagittal or axial plane at 12 μ m thickness on a cryostat (CM3050 S; Leica Microsystems, Wetzlar, Germany). The injured spinal cord sections were histologically evaluated by HE staining and immunohistochemistry, followed by quantitative analysis. The sections of transplanted spinal cord were subjected to HE staining, LFB staining, and immunohistochemistry followed by quantitative analyses.

For immunohistochemistry, tissue sections were stained with the following primary antibodies: anti-GFP (rabbit IgG, 1:200; Frontier Institute, Hokkaido, Japan), anti-Hu (human IgG, 1:1000, a gift from Dr. Robert Darnell, The Rockefeller University, New York, NY, USA), anti-GFAP (rat IgG, 1:200; Invitrogen), anti-APC CC-1 (mouse IgG, 1:200; Calbiochem, San Diego, CA, USA), anti-nestin (mouse IgG1, 1:500; BD Bioscience Pharmingen, San Jose, California, USA), anti-CS56 (a marker for CSPG, mouse IgM, 1:200; Sigma-Aldrich), anti-Iba1 (a marker for microglia/macrophages, rabbit IgG, 1:200; Wako Pure Chemical

Industries, Osaka, Japan), anti-arginase-1 (a marker for M2 macrophages, goat IgG, 1:200; Santa Cruz Biotechnology, Santa Cruz, CA, USA), anti-LAMP2 (a marker for endosomes or lysosomes, rat IgG, 1:200; Abcam, Cambridge, UK), anti-NF-H (mouse IgG1, 1:200; Chemicon, Millipore, Billerica, MA, USA), and anti-5HT (goat IgG, 1:200; Immunostar, Hudson, WI, USA). For immunohistochemistry with anti-GFP, -NF-H, and -5HT, a biotinylated secondary antibody (Jackson ImmunoResearch Laboratory Inc., West Grove, PA, USA) was used after exposing the sections to 0.3% H₂O₂ for 30 min at room temperature to inactivate endogenous peroxidases. The signals were enhanced with the Vectastain ABC kit (Vector Laboratories, Inc., Burlingame, CA, USA). Nuclei were stained with Hoechst 33258 (10 μ g/ml, Sigma-Aldrich). All images were obtained using a fluorescence microscope (BZ 9000; Keyence Co.) or a confocal laser scanning microscope (LSM 700; Carl Zeiss, Munich, Germany).

Quantitative analyses

Quantitative analyses of the histological findings (HE, LFB staining, and immunostaining for CS56, Iba1, NF-H, 5HT, GFP/each phenotypic marker, and arginase-1/Iba1) were performed using a BZ 9000 microscope and Dynamic Cell Count BZ-HIC software (Keyence Co.). The threshold values were maintained at a constant level for all analyses. The GFP⁺ area was quantified using images of axial sections of the lesion epicenter and 0.5 mm, 1.0 mm, 2.0 mm, 3.0 mm, and 4.0 mm rostral and caudal to the epicenter at 100 \times magnification (n = 4 each). To determine the spinal cord area, HE-stained images of axial sections of the lesion epicenter and 4.0 mm rostral and caudal to the epicenter at 100 \times magnification were used (n = 3 each). Quantitative analysis of the LFB⁺ area was similarly performed using axial sections of the lesion epicenter and 4.0 mm rostral and caudal to the epicenter at 100 \times magnification (n = 3 each). CS56⁺ areas as well as Iba1⁺ areas were quantified in the midsagittal sections of injured spinal cords at 100 \times magnification (n = 3 each).

To quantify NF-H⁺ fibers, four regions were automatically captured within the midsagittal sections of the lesion epicenter and 4.0 mm rostral and caudal to the epicenter at 400 \times magnification (n = 3 each). To assess the 5HT⁺ fibers, five automatically captured regions within the axial sections were analyzed at the lumbar intumescence, which was 6–8 mm caudal to the lesion epicenter (n = 3 each).

To quantify the proportion of each cell phenotype among the grafted cells *in vivo*, five regions were captured within sagittal sections at 200 \times magnification using an LSM 700 confocal laser scanning microscope. GFP and phenotypic marker double-positive cells were counted in each section (n = 3 each). To quantify the infiltrated M2 macrophages in the injured spinal cord,

the number of Iba1 and arginase-1 double-positive cells was counted within five regions of sagittal sections of the lesion epicenter at 200× magnification with an LSM 700 confocal laser scanning microscope (n = 3 each).

Statistical analysis

All data are reported as the mean ± SEM. An unpaired two-tailed Student's *t* test was used to evaluate the differences between groups with respect to microarray gene expression profile, *in vivo* BLI analysis, *in vivo* differentiation assays, and analyses of the CS56⁺, Iba1⁺ and GFP⁺ areas, and arginase-1⁺/Iba1⁺ cells. One-way ANOVA followed by the Tukey-Kramer test for multiple comparisons was used in the analyses of the HE, LFB, NF-H⁺, and 5HT⁺ areas and the Rotarod and DigiGait results. Repeated-measures two-way ANOVA followed by the Tukey-Kramer test was used for the BMS analysis. *P* values < 0.05 were considered statistically significant.

Abbreviations

NS/PCs: Neural stem/progenitor cells; SCI: Spinal cord injury; DPI: Days post-injury; BLI: Bioluminescence imaging; TP group: Transplanted group; ffluc: Fluorescent protein-fused luciferase; Tuj-1: βIII tubulin; GFAP: Glial fibrillary acidic protein; CNPase: 2'3'-cyclic nucleotide 3'-phosphodiesterase; CSPG: Chondroitin sulfate proteoglycan; PCA: Principal component analysis; iNOS: Inducible nitric oxide synthase; APC: Adenomatous polyposis coli antigen; HE: Hematoxylin-eosin; PBS: Phosphate-buffered saline; NF-H: 200 kDa neurofilament; 5HT: 5-hydroxytryptamine; BMS: Basso Mouse Scale; MEP: Motor-evoked potential; LFB: Luxol Fast Blue; ChABC: Chondroitinase ABC.

Competing interests

The authors have declared that no competing interests exist.

Authors' contributions

SN, AY, OT, HO, and MN designed the research; SN, AY, HI, MT, and HE performed research; SN, AY, and HI analyzed the data; SN, HO, and MN wrote the paper; and MN and HO supervised all the experiments. All authors read and approved the final manuscript.

Acknowledgments

We appreciate the help of Dr. A. Iwanami, Dr. Y. Takahashi, Dr. M. Shinozaki, Dr. T. Konomi, Dr. R. Zhang, Dr. G. Itakura, Dr. S. Tashiro, Dr. S. Kawabata, Dr. Y. Nishiyama, and Dr. K. Hori, members of the spinal cord research team at the Department of Orthopaedic Surgery, Rehabilitation Medicine and Physiology, Keio University School of Medicine. We also thank Ms. T. Harada, Ms. S. Miyao, Ms. M. Mizutani, and Ms. H. Shimada for their assistance with the experiments and animal care.

This work was supported by grants from the Japan Science and Technology–California Institute for Regenerative Medicine collaborative program; Grants-in-Aid for Scientific Research from the Japan Society for the Promotion of Science (SPS) and the Ministry of Education, Culture, Sports, Science, and Technology of Japan (MEXT); the Project for Realization of Regenerative Medicine; Support for Core Institutes for iPS Cell Research from the MEXT; Keio Gijuku Academic Development Funds; by a Grant-in-Aid for the Global COE program from MEXT to Keio University; and by a Grant-in-Aid for Scientific Research on Innovative Areas (Comprehensive Brain Science Network) from the MEXT.

Author details

¹Department of Orthopaedic Surgery, 35 Shinanomachi, Shinjuku, Tokyo 160-8582, Japan. ²Department of Physiology, Keio University School of Medicine, 35 Shinanomachi, Shinjuku, Tokyo 160-8582, Japan. ³Department of Orthopaedic Surgery, Saitama Social Insurance Hospital, 4-9-3 Kitaurawa, Urawa, Saitama 330-0074, Japan. ⁴Department of Orthopaedic Surgery, Murayama National Hospital Organization Murayama Medical Center, 2-37-1

Gakuen, Musashimurayama, Tokyo 208-0011, Japan. ⁵Genomic Science Laboratories, Dainippon Sumitomo Pharma Co, Ltd., 2-6-8 Doshoumachi, Chuo, Osaka 541-0045, Japan.

Received: 27 November 2012 Accepted: 25 December 2012

Published: 8 January 2013

References

1. Ogawa Y, Sawamoto K, Miyata T, Miyao S, Watanabe M, Nakamura M, Bregman BS, Koike M, Uchiyama Y, Toyama Y, Okano H: **Transplantation of in vitro-expanded fetal neural progenitor cells results in neurogenesis and functional recovery after spinal cord contusion injury in adult rats.** *J Neurosci Res* 2002, **69**:925–933.
2. Cummings BJ, Uchida N, Tamaki SJ, Salazar DL, Hooshmand M, Summers R, Gage FH, Anderson AJ: **Human neural stem cells differentiate and promote locomotor recovery in spinal cord-injured mice.** *Proc Natl Acad Sci USA* 2005, **102**:14069–14074.
3. Hofstetter CP, Holmström NAV, Lijja JA, Schweinhardt P, Hao J, Spenger C, Wiesenfeld-Hallin Z, Kurpad SN, Frisén J, Olson L: **Allodynia limits the usefulness of intraspinal neural stem cell grafts; directed differentiation improves outcome.** *Nat Neurosci* 2005, **8**:346–353.
4. Okada S, Ishii K, Yamane J, Iwanami A, Ikegami T, Katoh H, Iwamoto Y, Nakamura M, Miyoshi H, Okano HJ, Contag CH, Toyama Y, Okano H: **In vivo imaging of engrafted neural stem cells: its application in evaluating the optimal timing of transplantation for spinal cord injury.** *FASEB J* 2005, **19**:1839–1841.
5. Cao QL, Zhang YP, Howard RM, Walters WM, Tsoulfas P, Whittemore SR: **Pluripotent stem cells engrafted into the normal or lesioned adult rat spinal cord are restricted to a glial lineage.** *Exp Neurol* 2001, **167**:48–58.
6. Abematsu M, Tsujimura K, Yamano M, Saito M, Kohno K, Kohyama J, Namihira M, Komiya S, Nakashima K: **Neurons derived from transplanted neural stem cells restore disrupted neuronal circuitry in a mouse model of spinal cord injury.** *J Clin Invest* 2010, **120**:3255–3266.
7. Iwanami A, Kaneko S, Nakamura M, Kanemura Y, Mori H, Kobayashi S, Yamasaki M, Momoshima S, Ishii H, Ando K, Tanioka Y, Tamaoki N, Nomura T, Toyama Y, Okano H: **Transplantation of human neural stem cells for spinal cord injury in primates.** *J Neurosci Res* 2005, **80**:182–190.
8. Salazar DL, Uchida N, Hamers FPT, Cummings BJ, Anderson AJ: **Human Neural Stem Cells Differentiate and promote locomotor recovery in an early chronic spinal cord injury NOD-scid mouse model.** *PLoS One* 2010, **5**:e12272.
9. Parr AM, Kulbatski I, Tator CH: **Transplantation of adult rat spinal cord stem/progenitor cells for spinal cord injury.** *J Neurotrauma* 2007, **24**:835–845.
10. Kusano K, Enomoto M, Hirai T, Tsoulfas P, Sotome S, Shinomiya K, Okawa A: **Transplanted neural progenitor cells expressing mutant NT3 promote myelination and partial hindlimb recovery in the chronic phase after spinal cord injury.** *Biochem Biophys Res Commun* 2010, **393**:812–817.
11. Cusimano M, Biziato D, Brambilla E, Donega M, Alfaro-Cervello C, Snider S, Salani G, Pucci F, Comi G, Garcia-Verdugo JM, De Palma M, Martino G, Pluchino S: **Transplanted neural stem/precursor cells instruct phagocytes and reduce secondary tissue damage in the injured spinal cord.** *Brain* 2012, **135**:447–460.
12. Karimi-Abdolrezaee S, Eftekharpour E, Wang J, Morshead CM, Fehlings MG: **Delayed transplantation of adult neural precursor cells promotes remyelination and functional neurological recovery after spinal cord injury.** *J Neurosci* 2006, **26**:3377–3389.
13. Contag CH, Bachmann MH: **Advances in in vivo bioluminescence imaging of gene expression.** *Annu Rev Biomed Eng* 2002, **4**:235–260.
14. Hara-Miyayachi C, Tsuji O, Hanyu A, Okada S, Yasuda A, Fukano T, Akazawa C, Nakamura M, Imamura T, Matsuzaki Y, Okano HJ, Miyawaki A, Okano H: **Bioluminescent system for dynamic imaging of cell and animal behavior.** *Biochem Biophys Res Commun* 2012, **419**:188–193.
15. Kanemura Y, Mori H, Kobayashi S, Islam O, Kodama E, Yamamoto A, Nakanishi Y, Arita N, Yamasaki M, Okano H, Hara M, Miyake J: **Evaluation of in vitro proliferative activity of human fetal neural stem/progenitor cells using indirect measurements of viable cells based on cellular metabolic activity.** *J Neurosci Res* 2002, **69**:869–879.
16. Takahashi Y, Tsuji O, Kumagai G, Hara CM, Okano HJ, Miyawaki A, Toyama Y, Okano H, Nakamura M: **Comparative study of methods for administering neural stem/progenitor cells to treat spinal cord injury in mice.** *Cell Transplant* 2011, **20**:727–739.

17. Yasuda A, Tsuji O, Shibata S, Nori S, Takano M, Kobayashi Y, Takahashi Y, Fujiyoshi K, Hara CM, Miyawaki A, Okano HJ, Toyama Y, Nakamura M, Okano H: **Significance of remyelination by neural stem/progenitor cells transplanted into the injured spinal cord.** *Stem Cells* 2011, **29**:1983–1994.
18. Anderson AJ, Haus DL, Hooshmand MJ, Perez H, Sontag CJ, Cummings BJ: **Achieving stable human stem cell engraftment and survival in the CNS: is the future of regenerative medicine immunodeficient?** *Regen med* 2011, **6**:367–406.
19. Kumamaru H, Ohkawa Y, Saiwai H, Yamada H, Kubota K, Kobayakawa K, Akashi K, Okano H, Iwamoto Y, Okada S: **Direct isolation and RNA-seq reveal environment-dependent properties of engrafted neural stem/progenitor cells.** *Nat Commun* 2012, **3**:1140.
20. Nakamura M, Houghtling RA, MacArthur L, Bayer BM, Bregman BS: **Differences in cytokine gene expression profile between acute and secondary injury in adult rat spinal cord.** *Exp Neurol* 2003, **184**:313–325.
21. Kumamaru H, Saiwai H, Ohkawa Y, Yamada H, Iwamoto Y, Okada S: **Age-related differences in cellular and molecular profiles of inflammatory responses after spinal cord injury.** *J Cell Physiol* 2012, **227**:1335–1346.
22. Kumar S, Kahn MA, Dinh L, de Vellis J: **NT-3-mediated TrkC receptor activation promotes proliferation and cell survival of rodent progenitor oligodendrocyte cells in vitro and in vivo.** *J Neurosci Res* 1998, **54**:754–765.
23. Kokuzawa J, Yoshimura S, Kitajima H, Shinoda J, Kaku Y, Iwama T, Morishita R, Shimazaki T, Okano H, Kunisada T, Sakai N: **Hepatocyte growth factor promotes proliferation and neuronal differentiation of neural stem cells from mouse embryos.** *Mol Cell Neurosci* 2003, **24**:190–197.
24. Erlandsson A, Enarsson M, Forsberg-Nilsson K: **Immature neurons from CNS stem cells proliferate in response to platelet-derived growth factor.** *J Neurosci* 2001, **21**:3483–3491.
25. Johe KK, Hazel TG, Muller T, Dugich-Djordjevic MM, McKay RD: **Single factors direct the differentiation of stem cells from the fetal and adult central nervous system.** *Genes Dev* 1996, **10**:3129–3140.
26. Nori S, Okada Y, Yasuda A, Tsuji O, Takahashi Y, Kobayashi Y, Fujiyoshi K, Koike M, Uchiyama Y, Ikeda E, Toyama Y, Yamanaka S, Nakamura M, Okano H: **Grafted human-induced pluripotent stem-cell-derived neurospheres promote motor functional recovery after spinal cord injury in mice.** *Proc Natl Acad Sci USA* 2011, **108**:16825–16830.
27. Kitamura K, Iwanami A, Nakamura M, Yamane J, Watanabe K, Suzuki Y, Miyazawa D, Shibata S, Funakoshi H, Miyatake S, Coffin RS, Nakamura T, Toyama Y, Okano H: **Hepatocyte growth factor promotes endogenous repair and functional recovery after spinal cord injury.** *J Neurosci Res* 2007, **85**:2332–2342.
28. Hawryluk GW, Mothe A, Wang J, Wang S, Tator C, Fehlings MG: **An in vivo characterization of trophic factor production following neural precursor cell or bone marrow stromal cell transplantation for spinal cord injury.** *Stem Cell Dev* 2012, **21**:2222–2238.
29. Tuszyński MH, Gabriel K, Gage FH, Suhr S, Meyer S, Rosetti A: **Nerve growth factor delivery by gene transfer induces differential outgrowth of sensory, motor, and noradrenergic neurites after adult spinal cord injury.** *Exp Neurol* 1996, **137**:157–173.
30. Kumagai G, Okada Y, Yamane J, Nagoshi N, Kitamura K, Mukaino M, Tsuji O, Fujiyoshi K, Katoh H, Okada S, Shibata S, Matsuzaki Y, Toh S, Toyama Y, Nakamura M, Okano H: **Roles of ES cell-derived gliogenic neural stem/progenitor cells in functional recovery after spinal cord injury.** *PLoS One* 2009, **4**:e7706.
31. Bregman BS: **Spinal cord transplants permit the growth of serotonergic axons across the site of neonatal spinal cord transection.** *Brain Res* 1987, **431**:265–279.
32. Saruhashi Y, Young W, Perkins R: **The recovery of 5-HT immunoreactivity in lumbosacral spinal cord and locomotor function after thoracic hemisection.** *Exp Neurol* 1996, **139**:203–213.
33. Predy R, Malhotra SK: **Reactive astrocytes in lesioned rat spinal cord: effect of neural transplants.** *Brain Res Bull* 1989, **22**:81–87.
34. Grijalva I, Guizar-Sahagun G, Salgado-Ceballos H, Ibarra A, Franco-Bourland R, Espitia L, Madrazo I: **Improvement of host-graft adhesion by enzymatic manipulation of the subacute spinal cord contusion area in the rat.** *Transplant Proc* 1996, **28**:3340–3342.
35. Beck KD, Nguyen HX, Galvan MD, Salazar DL, Woodruff TM, Anderson AJ: **Quantitative analysis of cellular inflammation after traumatic spinal cord injury: evidence for a multiphasic inflammatory response in the acute to chronic environment.** *Brain* 2010, **133**:433–447.
36. Popovich PG, Wei P, Stokes BT: **Cellular inflammatory response after spinal cord injury in Sprague–Dawley and Lewis rats.** *J Comp Neurol* 1997, **377**:443–464.
37. Kigerl KA, Gensel JC, Ankeny DP, Alexander JK, Donnelly DJ, Popovich PG: **Identification of Two Distinct Macrophage Subsets with Divergent Effects Causing either Neurotoxicity or Regeneration in the Injured Mouse Spinal Cord.** *J Neurosci* 2009, **29**:13435–13444.
38. Block ML, Zecca L, Hong JS: **Microglia-mediated neurotoxicity: uncovering the molecular mechanisms.** *Nat Rev Neurosci* 2007, **8**:57–69.
39. Busch SA, Hamilton JA, Horn KP, Cuascut FX, Cutrone R, Lehman N, Deans RJ, Ting AE, Mays RW, Silver J: **Multipotent adult progenitor cells prevent macrophage-mediated axonal dieback and promote regrowth after spinal cord injury.** *J Neurosci* 2011, **31**:944–953.
40. Mukaino M, Nakamura M, Yamada O, Okada S, Morikawa S, Renault-Mihara F, Iwanami A, Ikegami T, Ohsugi Y, Tsuji O, Katoh H, Matsuzaki Y, Toyama Y, Liu M, Okano H: **Anti-IL-6-receptor antibody promotes repair of spinal cord injury by inducing microglia-dominant inflammation.** *Exp Neurol* 2010, **224**:403–414.
41. Schilling M, Besselmann M, Muller M, Strecker JK, Ringelstein EB, Kiefer R: **Predominant phagocytic activity of resident microglia over hematogenous macrophages following transient focal cerebral ischemia: an investigation using green fluorescent protein transgenic bone marrow chimeric mice.** *Exp Neurol* 2005, **196**:290–297.
42. Mantovani A, Sozzani S, Locati M, Allavena P, Sica A: **Macrophage polarization: tumor-associated macrophages as a paradigm for polarized M2 mononuclear phagocytes.** *Trends Immunol* 2002, **23**:549–555.
43. Busch SA, Horn KP, Silver DJ, Silver J: **Overcoming macrophage-mediated axonal dieback following CNS injury.** *J Neurosci* 2009, **29**:9967–9976.
44. Kokaia Z, Martino G, Schwartz M, Lindvall O: **Cross-talk between neural stem cells and immune cells: the key to better brain repair?** *Nat Neurosci* 2012, **15**:1078–1087.
45. Ziv Y, Avidan H, Pluchino S, Martino G, Schwartz M: **Synergy between immune cells and adult neural stem/progenitor cells promotes functional recovery from spinal cord injury.** *Proc Natl Acad Sci USA* 2006, **103**:13174–13179.
46. Andrews T, Zhang P, Bhat NR: **TNFalpha potentiates IFNgamma-induced cell death in oligodendrocyte progenitors.** *J Neurosci Res* 1998, **54**:574–583.
47. Ben-Hur T, Ben-Menachem O, Furer V, Einstein O, Mizrahi-Kol R, Grigoriadis N: **Effects of proinflammatory cytokines on the growth, fate, and motility of multipotential neural precursor cells.** *Mol Cell Neurosci* 2003, **24**:623–631.
48. Silver J, Miller JH: **Regeneration beyond the glial scar.** *Nat Rev Neurosci* 2004, **5**:146–156.
49. Ikegami T, Nakamura M, Yamane J, Katoh H, Okada S, Iwanami A, Watanabe K, Ishii K, Kato F, Fujita H, Takahashi T, Okano HJ, Toyama Y, Okano H: **Chondroitinase ABC combined with neural stem/progenitor cell transplantation enhances graft cell migration and outgrowth of growth-associated protein-43-positive fibers after rat spinal cord injury.** *Eur J Neurosci* 2005, **22**:3036–3046.
50. Bradbury EJ, Moon LD, Popat RJ, King VR, Bennett GS, Patel PN, Fawcett JW, McMahon SB: **Chondroitinase ABC promotes functional recovery after spinal cord injury.** *Nature* 2002, **416**:636–640.
51. Karimi-Abdolrezaee S, Eftekharpour E, Wang J, Schut D, Fehlings MG: **Synergistic Effects of Transplanted Adult Neural Stem/Progenitor Cells, Chondroitinase, and Growth Factors Promote Functional Repair and Plasticity of the Chronically Injured Spinal Cord.** *J Neurosci* 2010, **30**:1657–1676.
52. Reynolds BA, Tetzlaff W, Weiss S: **A multipotent EGF-responsive striatal embryonic progenitor cell produces neurons and astrocytes.** *J Neurosci* 1992, **12**:4565–4574.
53. Nagai T, Ibata K, Park ES, Kubota M, Mikoshiba K, Miyawaki A: **A variant of yellow fluorescent protein with fast and efficient maturation for cell-biological applications.** *Nat Biotechnol* 2002, **20**:87–90.
54. Yamane J, Nakamura M, Iwanami A, Sakaguchi M, Katoh H, Yamada M, Momoshima S, Miyao S, Ishii K, Tamaoki N, Nomura T, Okano HJ, Kanemura Y, Toyama Y, Okano H: **Transplantation of galectin-1-expressing human neural stem cells into the injured spinal cord of adult common marmosets.** *J Neurosci Res* 2010, **88**:1394–1405.
55. Lockhart DJ, Dong H, Byrne MC, Follettie MT, Gallo MV, Chee MS, Mittmann M, Wang C, Kobayashi M, Horton H, Brown EL: **Expression monitoring by hybridization to high-density oligonucleotide arrays.** *Nat Biotechnol* 1996, **14**:1675–1680.

56. Heishi M, Ichihara J, Teramoto R, Itakura Y, Hayashi K, Ishikawa H, Gomi H, Sakai J, Kanaoka M, Taiji M, Kimura T: **Global gene expression analysis in liver of obese diabetic db/db mice treated with metformin.** *Diabetologia* 2006, **49**:1647–1655.
57. Tsuji O, Miura K, Okada Y, Fujiyoshi K, Mukaino M, Nagoshi N, Kitamura K, Kumagai G, Nishino M, Tomisato S, Higashi H, Nagai T, Katoh H, Kohda K, Matsuzaki Y, Yuzaki M, Ikeda E, Toyama Y, Nakamura M, Yamanaka S, Okano H: **Therapeutic potential of appropriately evaluated safe-induced pluripotent stem cells for spinal cord injury.** *Proc Natl Acad Sci USA* 2010, **107**:12704–12709.
58. Basso DM, Fisher LC, Anderson AJ, Jakeman LB, McTigue DM, Popovich PG: **Basso Mouse Scale for locomotion detects differences in recovery after spinal cord injury in five common mouse strains.** *J Neurotraum* 2006, **23**:635–659.
59. Ogura H, Matsumoto M, Mikoshiba K: **Motor discoordination in mutant mice heterozygous for the type 1 inositol 1,4,5-trisphosphate receptor.** *Behav Brain Res* 2001, **122**:215–219.
60. Springer JE, Rao RR, Lim HR, Cho SI, Moon GJ, Lee HY, Park EJ, Noh JS, Gwag BJ: **The functional and neuroprotective actions of Neu 2000, a dual-acting pharmacological agent, in the treatment of acute spinal cord injury.** *J Neurotraum* 2010, **27**:139–149.

doi:10.1186/1756-6606-6-3

Cite this article as: Nishimura *et al.*: Time-dependent changes in the microenvironment of injured spinal cord affects the therapeutic potential of neural stem cell transplantation for spinal cord injury. *Molecular Brain* 2013 **6**:3.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

