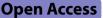
# **MICRO REPORT**



# Comprehensive identification of ubiquitin-like 3 (UBL3)-interacting proteins in the mouse brain

Hiroshi Ageta<sup>1\*</sup>, Tomoki Nishioka<sup>2,3</sup>, Hisateru Yamaguchi<sup>4</sup>, Kunihiro Tsuchida<sup>1\*</sup> and Natsumi Ageta-Ishihara<sup>5\*</sup>

# Abstract

Discovery of novel post-translational modifications provides new insights into changes in protein function, localization, and stability. They are also key elements in understanding disease mechanisms and developing therapeutic strategies. We have previously reported that ubiquitin-like 3 (UBL3) serves as a novel post-translational modifier that is highly expressed in the cerebral cortex and hippocampus, in addition to various other organs, and that 60% of proteins contained in small extracellular vesicles (sEVs), including exosomes, are influenced by UBL3. In this study, we generated transgenic mice expressing biotinylated UBL3 in the forebrain under control of the alpha-CaMKII promoter (Ubl3<sup>Tg/+</sup>). Western blot analysis revealed that the expression of UBL3 in the cerebral cortex and hippocampus was 6- to 7-fold higher than that in the cerebellum. Therefore, we performed immunoprecipitation of protein extracts from the cerebral cortex of  $Ubl3^{+/+}$  and  $Ubl3^{Tg/+}$  mice using avidin beads to comprehensively discover UBL3 interacting proteins, identifying 35 new UBL3 interacting proteins. Nine proteins were annotated as extracellular exosomes. Gene Ontology (GO) analysis suggested a new relationship between sEVs and RNA metabolism in neurodegenerative diseases. We confirmed the association of endogenous UBL3 with the RNAbinding proteins FUS and HPRT1—both listed in the Neurodegenerative Diseases Variation Database (NDDVD)and with LYPLA1, which is involved in Huntington's disease, using immunoprecipitation (IP)-western blotting analysis. These UBL3 interacting proteins will accelerate the continued elucidation of sEV research about proteins regulated by novel post-translational modifications by UBL3 in the brain.

**Keywords** Post-translational modification, Ubiquitin-like 3 (UBL3), Proteomics, Small extracellular vesicles (sEVs), RNA binding proteins, Neurodegenerative diseases

\*Correspondence: Hiroshi Ageta hiage@fujita-hu.ac.jp Kunihiro Tsuchida tsuchida@fujita-hu.ac.jp Natsumi Ageta-Ishihara natsumi.ageta-Ishihara@sci.toho-u.ac.jp <sup>1</sup>Division for Therapies Against Intractable Diseases, Center for Medical Science, Fujita Health University, 1-98 Dengakugakubo, Kutsukake-cho, Toyoake, Aichi 470-1192, Japan



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/. The Creative Commons Public Domain Dedication waiver (http://creativecommons.org/publicdomain/zero/1.0/) applies to the data made available in this article, unless otherwise stated in a credit line to the data.

 <sup>2</sup>Open Facility Center, Research Promotion Headquarters, Fujita Health University, Toyoake, Aichi 470-1192, Japan
<sup>3</sup>Division of Cell Biology, International Center for Brain Science, Fujita Health University, Toyoake, Aichi 470-1192, Japan
<sup>4</sup>Department of Medical Technology, Yokkaichi Nursing and Medical Care University, Yokkaichi 512-8045, Japan
<sup>5</sup>Department of Biomolecular Science, Faculty of Science, Toho University,

2-2-1 Miyama, Funabashi, Chiba 274-8510, Japan



# Main text

Post-translational modifications precisely regulate protein function and stability and play important roles in various cellular processes, including signal transduction [1, 2]. They are also very important in drug development because the precise control of target molecules involved in specific diseases and pathological conditions is an absolute requirement for designing effective therapies [3]. In the nervous system, post-translational modifications such as phosphorylation and ubiquitination are essential for the precise regulation of neurogenesis and synaptic function, and are also involved in the onset and progression of neurodegenerative diseases [4–6]. Therefore, posttranslational modifications play a crucial role in understanding disease mechanisms and developing therapeutic strategies.

We previously reported that ubiquitin-like 3 (UBL3)/ membrane-anchored ubiquitin-fold protein is a novel post-translational modifier and that UBL3 regulates the transport of 60% of proteins contained in sEVs including exosomes [2, 7]. Furthermore, we comprehensively identified UBL3-associated molecules in MDA-MB-231 human breast cancer cells. Among these proteins, the oncogenic protein Ras undergoes UBL3 modification, is encapsulated in sEVs, is taken up by other cells, and activates growth signaling [7]. We have also reported that UBL3-interacting proteins identified in MDA-MB-231 cells include neurodegenerative disease-related proteins such as presenilin 1 and huntingtin-interacting protein 1-related protein. Although UBL3 is highly expressed in the brain [7], UBL3-associated molecules have not yet been identified in the brain. Therefore, we generated transgenic mice expressing UBL3 in the forebrain in this study and comprehensively identified UBL3-interacting proteins in the brain.

Because UBL3 mRNA has been detected in neurons in the Allen Brain Atlas, we constructed a transcription unit by inserting an artificially synthesized coding region for mouse Ubl3 with a biotinylated tag added to the N-terminus into the alpha-CaMKII promoter. Sfi I fragments (Fig. 1a) were microinjected into the pronuclei of one-cell embryos of C57BL/6J mice to produce transgenic mice using previously reported methods [8]. Microinjected embryos were transferred to the oviducts of pseudopregnant females. Founder transgenic mice were identified by PCR using genomic DNA extracted from their tails (Fig. 1b) and bred with C57BL/6J mice (CLEA). Between *Ubl3*<sup>+/+</sup> (wild type; WT) and *Ubl3*<sup>Tg/+</sup> (transgenic; TG) male mice (9-10-week-old), no differences were observed for body (Fig. 1c) or brain weight (Fig. 1d). As we previously confirmed endogenous UBL3 expression in the cerebral cortex, hippocampus, and cerebellum using western blotting [7], we measured the respective weights of these tissues in Ubl3<sup>+/+</sup> and Ubl3<sup>Tg/+</sup> male mice. We did not detect statistically significant differences in the weight of the cortex (Fig. 1e), hippocampus (Fig. 1f), or cerebellum (Fig. 1g). Therefore, we quantified the amount of biotinylated UBL3 overexpressed in the cerebral cortex, hippocampus, and cerebellum. The amount of overexpressed UBL3 detected with a streptavidin-HRP (Invitrogen, 19534-050, 1:5000) was normalized to the amount of endogenous control protein detected with a glyceraldehyde 3-phosphate dehydrogenase (GAPDH) antibody (Cell Signaling, 2118, 1:1000). The results showed that UBL3 expression was significantly higher in the cerebral cortex and hippocampus than in the cerebellum (Fig. 1h, i). Because sufficient protein content was extracted from the cortex compared with the hippocampus, each protein extract from the cortex of *Ubl3*<sup>+/+</sup> and *Ubl3*<sup>Tg/+</sup> male mice (9-10-week-old, n=3in each group) was incubated with 30 µL of NeutrAvidin Agarose (ThermoFisher, 29202) for 18 h at 4 °C and subjected the proteins to mass spectrometry using previously reported methods [7]. The peptides were analyzed by liquid chromatography/mass spectrometry (LC/MS) using an Orbitrap Fusion mass spectrometer coupled to an Easy-nLC 1000 using an EASY-Spray ES900 column system (ThermoFisher, 75  $\mu$ m × 150 mm, particle size 3  $\mu$ m). Mass spectrometry raw files were processed using Proteome Discoverer version 2.4 (ThermoFisher) and a local MASCOT server (version 2.6.2; Matrix Science). The MS/MS data were searched against Mus musculus (SwissProt TaxID=10090\_and\_subtaxonomies) (v2017-10-25). We identified 35 UBL3 interacting molecules with an increase of 1.5 times or more in the *Ubl3*<sup>Tg/+</sup> mice compared to the *Ubl3*<sup>+/+</sup> mice and an experimental q-value of 0.05 or less (Table 1). When a protein-protein interaction network was created using the STRING database, 9 out of 35 UBL3-binding molecules were annotated as extracellular exosome (GO:0070062) (Fig. 1j). The percentage of molecules annotated as exosomes was 25%, which was similar to the comprehensive proteomic analysis of UBL3 interacting proteins conducted in breast cancer cells [7]. The ClusterProfiler R package was used for GO enrichment analysis of these UBL3 interacting proteins, which categorized these molecules as RNA binding proteins (Fig. 1k, l). Recent studies have focused on the relationship between abnormalities in RNA metabolism caused by the disruption of RNA binding proteins and neurodegenerative diseases [9]. Interestingly, the UBL3 interacting proteins identified in this study included the RNA binding proteins FUS, Hnrnpa1, and Hprt1, which are included in the 289 genes registered in the Neurodegenerative Diseases Variation Database (NDDVD, http:// www.sysbio.org.cn/NDDVD/diseases). Protein extract from the cortex of  $Ubl3^{+/+}$  male mice (9-week-old, n=3in each group) were incubated with 25 µL of Protein G Sepharose (Cytiva, 17-0618-01) and 2 µg of normal

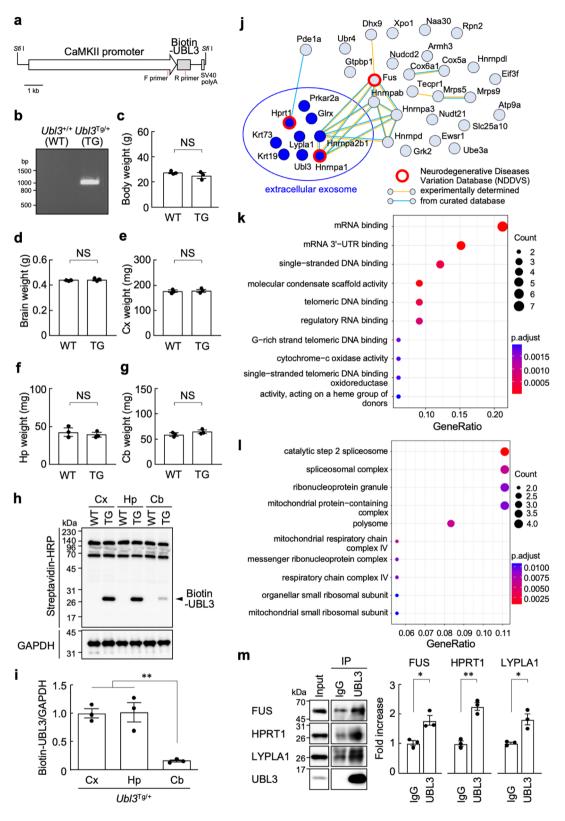


Fig. 1 (See legend on next page.)

# (See figure on previous page.)

**Fig. 1** UBL3-interacting molecules identified from the cerebral cortex in transgenic mice overexpressing UBL3 under the control of the alpha-CaMKII promoter. **a**, Schematic representation of the UBL3 transgenic construct. Red lines indicate the location of the PCR primers used for genotyping. **b**, Representative agarose gel image of the RT-PCR genotyping.  $Ubl3^{Tg/+}$  mice showed an amplified transgene of Ubl3 as 931 bp. Forward primer, 5'-CTTTCTCA AGGACCATCCCA-3'. Reverse primer, 5'-GCTGCTGACCTGCTCTTT-3'. **c**, Body weights of  $Ubl3^{+/+}$  and  $Ubl3^{Tg/+}$  male mice (9-10-week-old). **d**, Whole brain weights of  $Ubl3^{+/+}$  and  $Ubl3^{Tg/+}$  male mice (9-10-week-old). **e**, Cortex weight. **f**, Hippocampus weight. **g**, Cerebellum weight. **h**, **i**, Western blot analysis of the levels of biotinylated UBL3 and GAPDH (internal control) in proteins extracted from the cortex (Cx), hippocampus (Hp), and cerebellum (Cb) of  $Ubl3^{+/+}$  and  $Ubl3^{Tg/+}$  male mice (9-10-week-old). **h**. Quantitative analysis of the normalized fraction of biotinylated UBL3 (**i**). **j**, Protein-protein interaction network of UBL3-interacting molecules in the mouse cortex obtained from the STRING database. The protein group surrounded by a blue circle was annotated as an extracellular exosome protein (GO:0070062). **k**, GO analysis of molecular functions of UBL3-interacting molecules. **l**, GO analysis for the cellular components of UBL3-interacting molecules. **m**, IP/western blot assay of UBL3 either with FUS, HPRT1, or LYPLA1 from  $Ubl3^{+/+}$  (9 week-old male mice) cerebral cortex lysates. Densitometric quantification of the relative amount of each protein. Data are mean ± s.e.m. **c-g**, **m**, Two-tailed unpaired *t* test. **i**, One-way ANOVA with Tukey's multiple comparison test. \**P* < 0.05, \*\**P* < 0.01, NS, not significant

	Accession No.	Description	# AAs	Gene Symbol	extracel- lular exosome	Neurodegen- erative Diseas- es Variation Database
1	P12787	Cytochrome c oxidase subunit 5 A, mitochondrial [OS=Mus musculus]	146	Cox5a	-	-
2	Q61545	RNA-binding protein EWS [OS=Mus musculus]	655	Ewsr1	-	-
3	P56959	RNA-binding protein FUS [OS=Mus musculus]	518	Fus	-	+
4	Q99MK8	Beta-adrenergic receptor kinase 1 [OS=Mus musculus]	689	Grk2	-	-
5	O08582	GTP-binding protein 1 [OS=Mus musculus]	668	Gtpbp1	-	-
6	P19001	Keratin, type I cytoskeletal 19 [OS=Mus musculus]	403	Krt19	+	-
7	Q6NXH9	Keratin, type II cytoskeletal 73 [OS=Mus musculus]	539	Krt73	+	-
3	P97823-1	Acyl-protein thioesterase 1 [OS=Mus musculus]	230	Lypla1	+	-
9	Q9D7N3	28 S ribosomal protein S9, mitochondrial [OS=Mus musculus]	390	Mrps9	-	-
10	Q9Z2M6	Ubiquitin-like protein 3 [OS=Mus musculus]	117	Ubl3	+	-
11	A2AN08	e3 ubiquitin-protein ligase UBR4 [OS=Mus musculus]	5180	Ubr4	-	-
12	Q99020	Heterogeneous nuclear ribonucleoprotein A/B [OS=Mus musculus]	285	Hnrnpab	-	-
13	Q9Z130	Heterogeneous nuclear ribonucleoprotein D-like [OS=Mus musculus]	301	Hnrnpdl	-	-
14	Q6PD19-1	UPF0668 protein C10orf76 homolog [OS=Mus musculus]	689	Armh3	-	-
15	O70133-2	Isoform 2 of ATP-dependent RNA helicase A [OS=Mus musculus]	1381	Dhx9	-	-
16	Q9QZD8	Mitochondrial dicarboxylate carrier [OS=Mus musculus]	287	Slc25a10	-	-
7	P49312-1	Heterogeneous nuclear ribonucleoprotein A1 [OS=Mus musculus]	320	Hnrnpa1	+	+
8	Q99N87	28 S ribosomal protein S5, mitochondrial [OS=Mus musculus]	432	Mrps5	-	-
9	070228	Probable phospholipid-transporting ATPase IIA [OS=Mus musculus]	1047	Atp9a	-	-
20	P00493	Hypoxanthine-guanine phosphoribosyltransferase [OS=Mus musculus]	218	Hprt1	+	+
21	Q9QUH0	Glutaredoxin-1 [OS=Mus musculus]	107	Glrx	+	-
22	O88569	heterogeneous nuclear ribonucleoproteins A2/B1 [OS=Mus musculus]	353	Hnrnpa2b1	+	-
23	Q9DCH4	Eukaryotic translation initiation factor 3 subunit F [OS=Mus musculus]	361	Eif3f	-	-
24	Q61481	Dual specificity calcium/calmodulin-dependent 3',5'-cyclic nucleotide phosphodiesterase 1 A OS=Mus musculus OX=10,090 GN=Pde1a PE=1 SV=3	545	Pde1a	-	-
25	Q8CES0-1	N-alpha-acetyltransferase 30 [OS=Mus musculus]	364	Naa30	-	-
26	P43024	Cytochrome c oxidase subunit 6A1, mitochondrial [OS=Mus musculus]	111	Cox6a1	-	-
27	Q9DBG6	Dolichyl-diphosphooligosaccharideprotein glycosyltransferase subunit 2 [OS=Mus musculus]	631	Rpn2	-	-
28	Q9CQ48	NudC domain-containing protein 2 [OS=Mus musculus]	157	Nudcd2	-	-
29	P12367	cAMP-dependent protein kinase type II-alpha regulatory subunit [OS=Mus musculus]	401	Prkar2a	+	-
30	Q9CQF3	Cleavage and polyadenylation specificity factor subunit 5 [OS=Mus musculus]	227	Nudt21	-	-
31	O08759	Ubiquitin-protein ligase E3A [OS=Mus musculus]	870	Ube3a	-	-
32	Q80VP0-1	Tectonin beta-propeller repeat-containing protein 1 [OS=Mus musculus]	1166	Tecpr1	-	-
33	Q6P5F9	Exportin-1 [OS=Mus musculus]	1071	Xpo1	-	-
34	Q60668-1	heterogeneous nuclear ribonucleoprotein D0 [OS=Mus musculus]	355	Hnrnpd	-	-
35	Q8BG05	Heterogeneous nuclear ribonucleoprotein A3 [OS=Mus musculus]	379	Hnrnpa3	-	-

Table 1 T	he list of 35	UBL3 interacting	proteins
-----------	---------------	------------------	----------

rabbit immunoglobulin G (IgG) (Cell Signaling, 2729 S) or anti-UBL3 antibody (Proteintech, 14100-1-AP, lot 00005139) for 18 h at 4 °C and subjected western blotting either with anti-FUS (Santa Cruz, sc-47711, 1:200) or anti- HPRT1 (abcam, ab109021, 1:20000) antibodies. As a result, we showed that endogenous UBL3 is associated with FUS and HPRT1 (Fig. 1m). TDP-43, which is related to neurodegenerative diseases, has been reported to be encapsulated in sEVs [10] and binds to FUS [11]. Whether the transport of TDP-43 to sEVs is mediated by the UBL3 modification of FUS is a subject for future studies. If the relationship between RNA metabolism and protein transport to sEVs in neurodegenerative diseases is elucidated, compounds that affect UBL3 modification may become novel drug candidates, employing innovative therapeutic strategies for neurodegenerative diseases involving RNA metabolism. We previously reported that huntingtin-interacting protein 1-related protein associates with UBL3 in MDA-MB-231 cells [7]. Interestingly, among the UBL3 interacting molecules in the cerebral cortex, we identified Lypla1/APT1, a molecule involved in the pathogenesis of Huntington's disease [12], and confirmed its binding to UBL3 using immunoprecipitation (IP)-western blotting with anti-LYPLA1 antibody (Proteintech, 16055-1-AP, 1:2000) (Fig. 1m). This result suggests that UBL3 could be involved in the pathogenesis and/or progression of neurodegenerative diseases.

In this study, we successfully established transgenic mice that highly expressed biotinylated UBL3, specifically in the forebrain. Beta-amyloid in Alzheimer's disease and alpha-synuclein in Parkinson's disease are known to be sorted through sEVs [13, 14]. We have previously reported that 60% of proteins sorted into sEVs are UBL3-dependent [7]. sEVs research in the field of neuroscience is expected to be accelerated by crossbreeding mouse models of sEV-associated neurodegenerative diseases with the newly established mice overexpressing UBL3 in the forebrain.

#### Abbreviations

UBL3	Ubiquitin-like 3
sEVs	Small extracellular vesicles
GO	Gene Ontology
WT	Wild type
TG	Transgenic
GAPDH	Glyceraldehyde 3-phosphate dehydrogenase
NDDVD	Neurodegenerative Diseases Variation Database
lgG	Immunoglobulin G
IP	Immunoprecipitation

#### Acknowledgements

We thank C. Ohshima and M. Natsume (Fujita Health University) for the technical help; Dr. M. Mayford (University of California) for pMM403 vector containing the alpha-CaMKII promoter.

#### Authors' contributions

H.A. and N.A.-I. performed experiments. T. N. and H. Y. performed proteomics analyses. K.T. supervised the manuscript. H.A. and N.A.-I. designed experiments and wrote the manuscript. All authors read and approved the final manuscript.

#### Funding

This work was supported by JSPS KAKENHI Grant Number JP21K07159 and 21H00293 (to H.A.) and 23K06394 (to N.A.-I.), AMED Grant Number AMED 22wm0525011h0002 and 23wm0525011h0002 (to N.A.-I.), and JST, PRESTO Grant Number JPMJPR21E1 (to N.A.-I.), by the research grants from TUGRIP (Toho University Grant for Research Initiative Program), Koyanagi-Foundation, Chugai Foundation for Innovative Drug Discovery Science, Research Foundation for Qbto-Science and Technology, and Astellas Foundation for Research on Metabolic Disorders (to N.A.-I.). This study is also supported by an Intramural Research Grant (5–6) for Neurological and Psychiatric Disorders of the NCNP and a grant from the Fujita Health University (to K.T.).

#### Data availability

All data analyzed during this study are included in this published article and its additional file. Proteomics raw datasets are deposited in jPOST (under accession codes JPST002396, PXD047087).

## Declarations

## Ethics approval and consent to participate

All animal care and procedures performed in this study were undertaken according to the rules and regulations of the Guide for the Care and Use of Laboratory Animals. The experimental protocols were approved by the Animal Care Committee of Laboratory Animals of the Fujita Health University (AP23003).

#### **Consent for publication**

Not applicable.

#### **Competing interests**

There are no competing interests to declare in relation to this manuscript.

Received: 30 November 2023 / Accepted: 7 August 2024 Published online: 15 August 2024

#### References

- Deribe YL, Pawson T, Dikic I. Post-translational modifications in signal integration. Nat Struct Mol Biol. 2010;17:666–72.
- Ageta H, Tsuchida K. Post-translational modification and protein sorting to small extracellular vesicles including exosomes by ubiquitin and UBLs. Cell Mol Life Sci. 2019;76:4829–48.
- Ahearn IM, Haigis K, Bar-Sagi D, Philips MR. Regulating the regulator: posttranslational modification of RAS. Nat Rev Mol Cell Biol. 2011;13:39–51.
- 4. Tai HC, Schuman EM. Ubiquitin, the proteasome and protein degradation in neuronal function and dysfunction. Nat Rev Neurosci. 2008;9:826–38.
- Ageta-Ishihara N, Takemoto-Kimura S, Kondo Y, Okamura M, Bito H. Lipidation states orchestrate CLICK-III/CaMKIgamma's stepwise association with Golgi and rafts-enriched membranes and specify its functional coupling to STEF-Rac1-dependent neurite extension. Front Cell Neurosci. 2023;17:1204302.
- Ageta-Ishihara N, Miyata T, Ohshima C, Watanabe M, Sato Y, Hamamura Y, Higashiyama T, Mazitschek R, Bito H, Kinoshita M. Septins promote dendrite and axon development by negatively regulating microtubule stability via HDAC6-mediated deacetylation. Nat Commun. 2013;4:2532.
- Ageta H, Ageta-Ishihara N, Hitachi K, Karayel O, Onouchi T, Yamaguchi H, Kahyo T, Hatanaka K, Ikegami K, Yoshioka Y, et al. UBL3 modification influences protein sorting to small extracellular vesicles. Nat Commun. 2018;9:3936.
- Ageta H, Murayama A, Migishima R, Kida S, Tsuchida K, Yokoyama M, Inokuchi K. Activin in the brain modulates anxiety-related behavior and adult neurogenesis. PLoS ONE. 2008;3:e1869.
- Nussbacher JK, Tabet R, Yeo GW, Lagier-Tourenne C. Disruption of RNA metabolism in neurological diseases and emerging therapeutic interventions. Neuron. 2019;102:294–320.
- Iguchi Y, Eid L, Parent M, Soucy G, Bareil C, Riku Y, Kawai K, Takagi S, Yoshida M, Katsuno M, et al. Exosome secretion is a key pathway for clearance of pathological TDP-43. Brain. 2016;139:3187–201.
- Kim SH, Shanware NP, Bowler MJ, Tibbetts RS. Amyotrophic lateral sclerosisassociated proteins TDP-43 and FUS/TLS function in a common biochemical complex to co-regulate HDAC6 mRNA. J Biol Chem. 2010;285:34097–105.

- Virlogeux A, Scaramuzzino C, Lenoir S, Carpentier R, Louessard M, Genoux A, Lino P, Hinckelmann MV, Perrier AL, Humbert S, Saudou F. Increasing brain palmitoylation rescues behavior and neuropathology in Huntington disease mice. Sci Adv 2021, 7.
- 13. Lee HJ, Bae EJ, Lee SJ. Extracellular alpha–synuclein-a novel and crucial factor in Lewy body diseases. Nat Rev Neurol. 2014;10:92–8.
- Rajendran L, Honsho M, Zahn TR, Keller P, Geiger KD, Verkade P, Simons K. Alzheimer's disease beta-amyloid peptides are released in association with exosomes. Proc Natl Acad Sci U S A. 2006;103:11172–7.

# **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.